Phoxim acts as an environmental stress for insect parasitoid wasp

Yan Song ¹, Feng-Ming Gu ¹, Jun Wang ^{1,2}, Fu-An Wu ^{1, 2}, Sheng Sheng ^{1,2,*}

¹ School of biotechnology, Jiangsu University of Science and Technology, 212100 Zhenjiang, China;

² Sericultural Research Institute, Chinese Academy of Agricultural Sciences, 212100 Zhenjiang, China.

Key words: Environmental stress; Insecticides; Parasitic wasps; Detoxifying enzymes; Behavioral ecology.

*Presenting author email: sheng@just.edu.cn

Chemical insecticides play essential roles in controlling the agricultural insect pests during the past decades. However, more evidence indicates that the insecticides impose great stresses such as environmental pollution, pest resistance and human health. Although a plenty of researches well demonstrated the mechanism of pest resistance under insecticide stress (Ehler, 2010), its impact on non-target insects still receives less attention. Insect parasitic wasps are often used as biocontrol agents in agroecosystems to manager the increase of insect pest's population. On the other hand, they can easily encounter insecticides through direct exposure to spray droplets or residues on crop foliage (Jepson, 1989; Desneux et al., 2004), resulting in the failure of pest control of parasitoid wasps. As the natural enemies and non-target insects, the detail information of how parasitic wasps response insecticides stress still remains unknown.

In the present study, we chose the insecticide O, O-diethyl O-(alpha-cyanobenzylideneamino) phosphorothioate (phoxim) (Wang et al., 2013) and parasitic wasp *Meteorus pulchricornis* (Sheng et al., 2014) as the research system, aiming to evaluate the effect of phoxim on behavioral response and detoxifying system in *M. pulchricornis*. The behavioral tests revealed that after exposure to LC_{30} phoxim, female *M. pulchricornis* adults significantly shortened their patch residence time (Fig. 1). Meanwhile, the oviposition behavior (the number of oviposition) was also negatively affected by LC_{30} phoxim. By using RNA-Seq technology, we screened hundreds of differentially expressed genes and among these, seven detoxifying enzyme genes, including cytochrome P450 and esterase genes, were identified (Table 1). Interestingly, all these seven genes were down-regulated, suggesting the LC_{30} phoxim can inhibit the transcriptional levels of main detoxifying enzyme genes in *M. pulchricornis*.

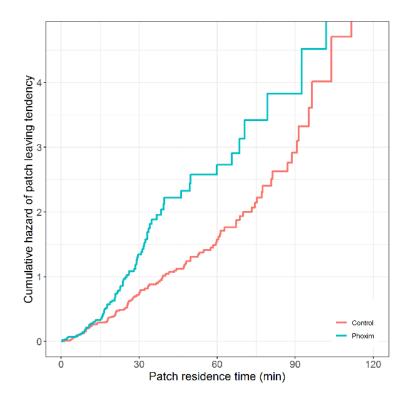


Fig. 1 Cumulative leaving tendency (hazard functions) of M. pulchricornis in response to phoxim

Gene ID	Log ₂ fold Change	P value	Gene Description	Up/Down regulation
Cytochrome P450				
Cluster-6774.4984	-1.47	4.53E-48	carboxylesterase-6-like	Down
Cluster-6774.5536	-1.42	8.03E-08	cytochrome P450 6k1-like	Down
Cluster-6774.5564	-1.84	1.61E-09	cytochrome P450 4g15	Down
Cluster-6774.6700	-1.67	8.85E-18	cytochrome P450 4g15	Down
Cluster-6774.8625	-1.22	3.93E-05	cytochrome P450 4C1-like	Down
Esterase				
Cluster-6774.5376	-1.80	2.74E-27	esterase FE4	Down
Cluster-6774.7298	-2.0	3.35 E-5	esterase E4	Down

Table 1 The differentially detoxifying enzyme expressed genes in *M. pulchricornis* after they exposed to LC_{30} phoxim

In conclusion, the present study demonstrated that the commonly used insecticide phoxim can act a strong environmental stress for the non-target and natural enemy parasitic wasp *M. pulchricornis* by lowering its behavioral efficacy and inhibiting the expression of detoxifying enzyme genes. Therefore, the application of phoxim must be cautious in the agroecosystem in the future.

Acknowledgements: This work was supported by the National Natural Science Foundation of China (31500312), the Key Research and Development Program (Modern Agriculture) of Zhenjiang City (NY2019021), and the Special Fund for China Agricultural Research System (CARS-18).

Reference

- [1] Ehler, L.E., 2010. Integrated pest management (IPM): definition, historical development and implementation, and the other IPM. Pest Manag. Sci. 62 (9), 787–789.
- [2] Jepson PC., 1989. The Temporal and Spatial Dynamics of Pesticide Side-effects on Non-target. Invertebrates. Intercept, Wimborne, UK: P. C. Jepson, pp. 95–127.
- [3] Desneux N, Wajnberg E, Fauvergue X, Privet S and Kaiser L.,2004. Oviposition behaviour and patch-time allocation in two aphid parasitoids exposed to deltamethrin residues. Entomologia Experimentalis et Applicata 112, 227–235.
- [4] Wang YH, Gu ZY, Wang JM, Sun SS, Wang BB, Jin YQ, Shen WD and Li B., 2013. Changes in. the activity and the expression of detoxification enzymes in silkworms (*Bombyx mori*) after phoxim feeding. Journal of Insect Behavior 105, 13–17.
- [5] Sheng S, Feng S, Meng L and Li B.,2014. Departure mechanisms for host search on high density. patches in the parasitoid *Meteorus pulchricornis* (Wesmael) (Hymenoptera: Braconidae). Journal of Insect Science 14, 1–5.