



Optimization of Pulsed Electric Fields-Assisted Extraction of phenolic compounds from white and red grape pomace using Response Surface Methodology

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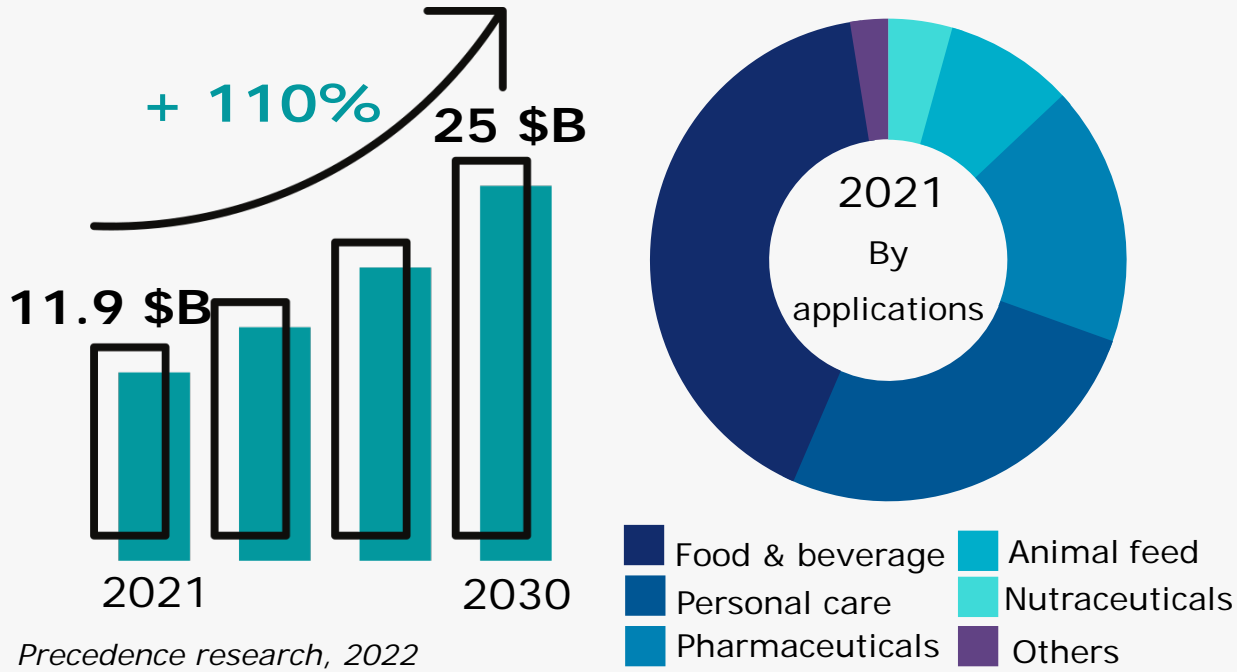
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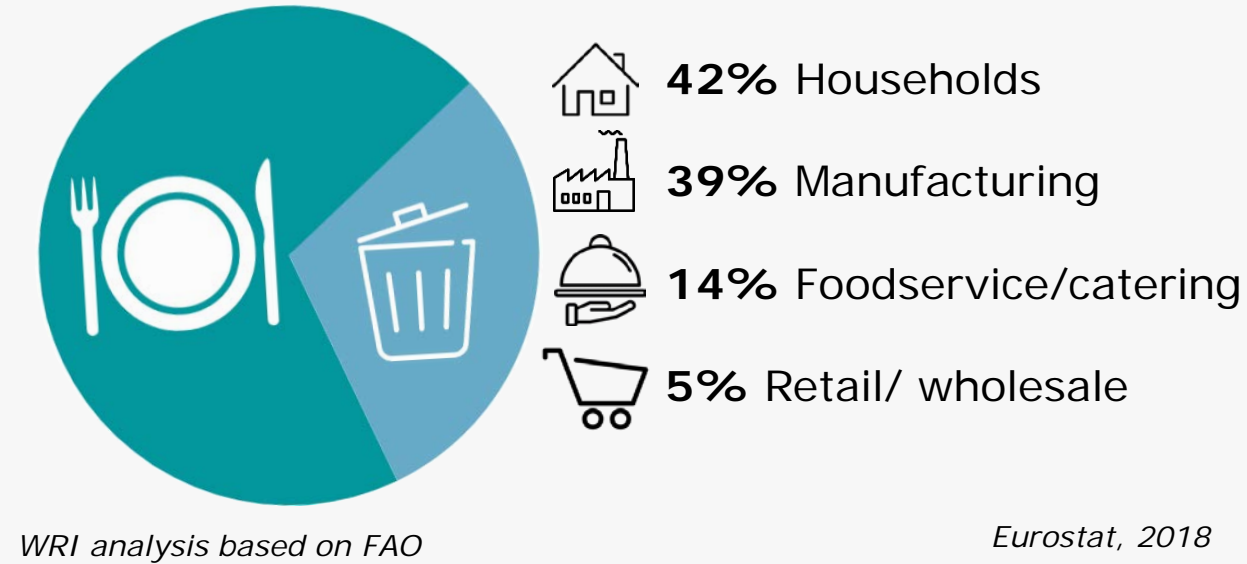


16/06/2022, Corfù, Greece

Global natural extracts market



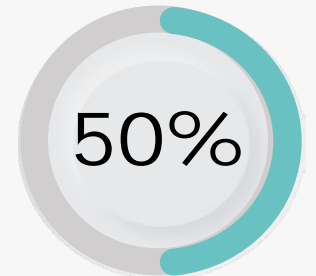
Food waste and by-products: a low-cost source of bioactive compounds



Intensive demand for natural ingredients



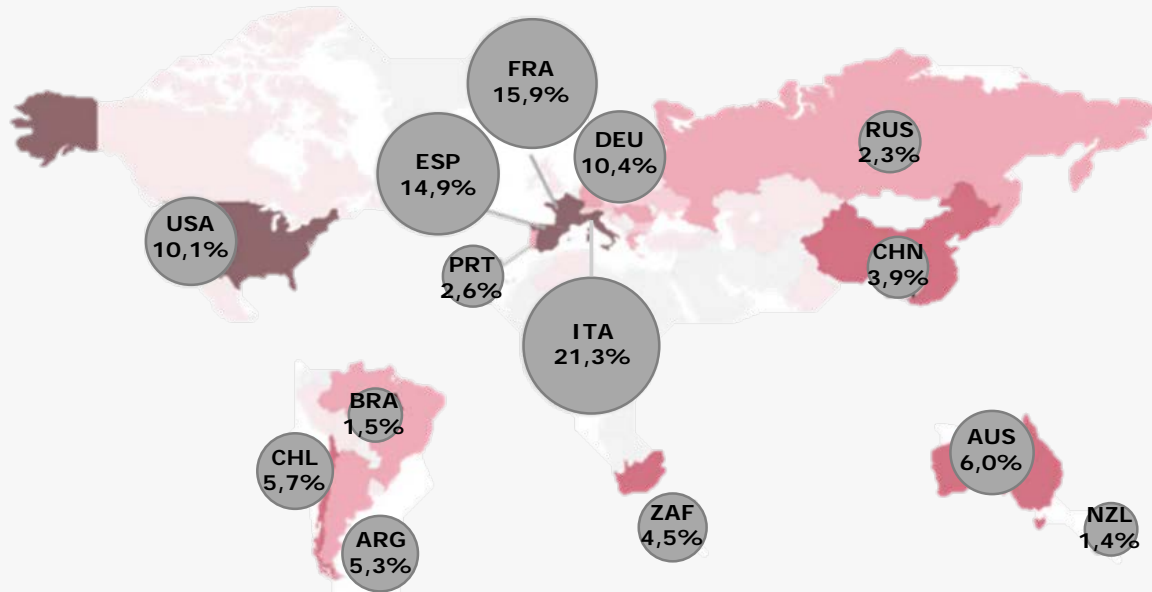
Innova Market Insights, Top Ten Trends for 2021



Call for a reduction of global food waste by 2030

Wine production in 2021

236 MhL of wine produced in 2021

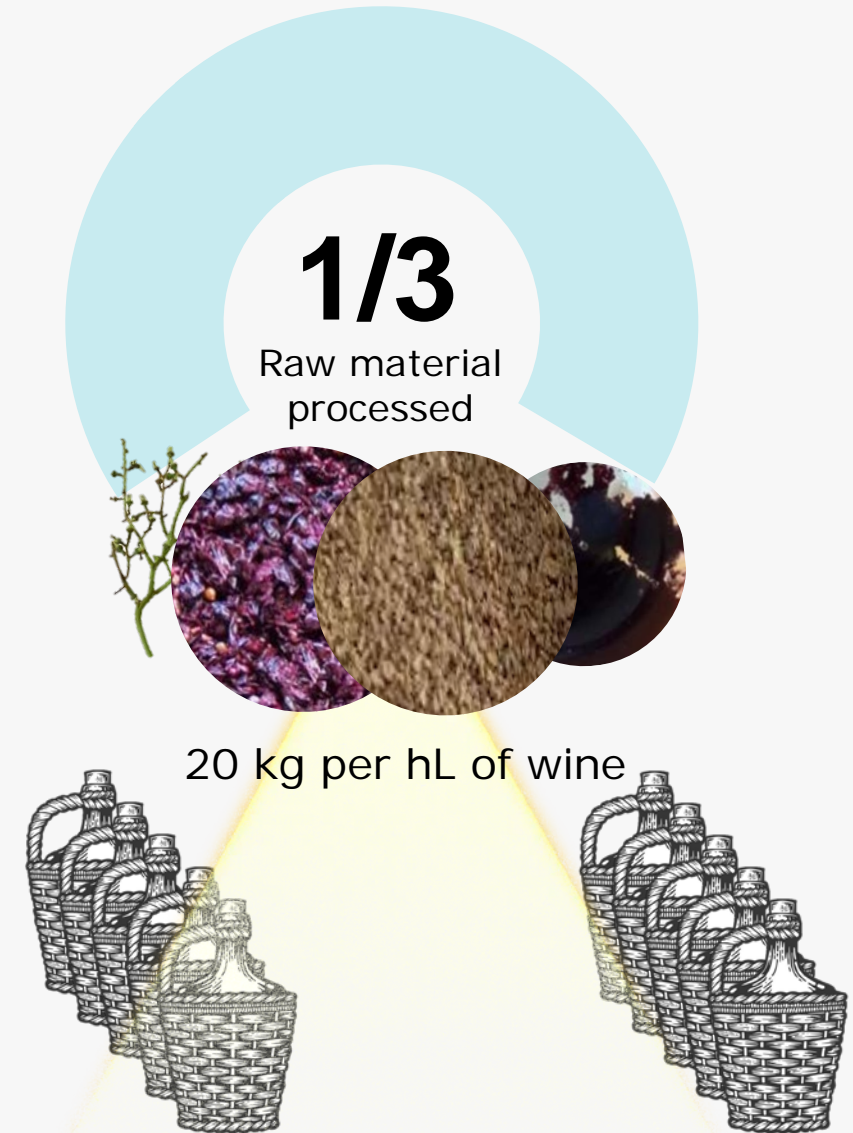


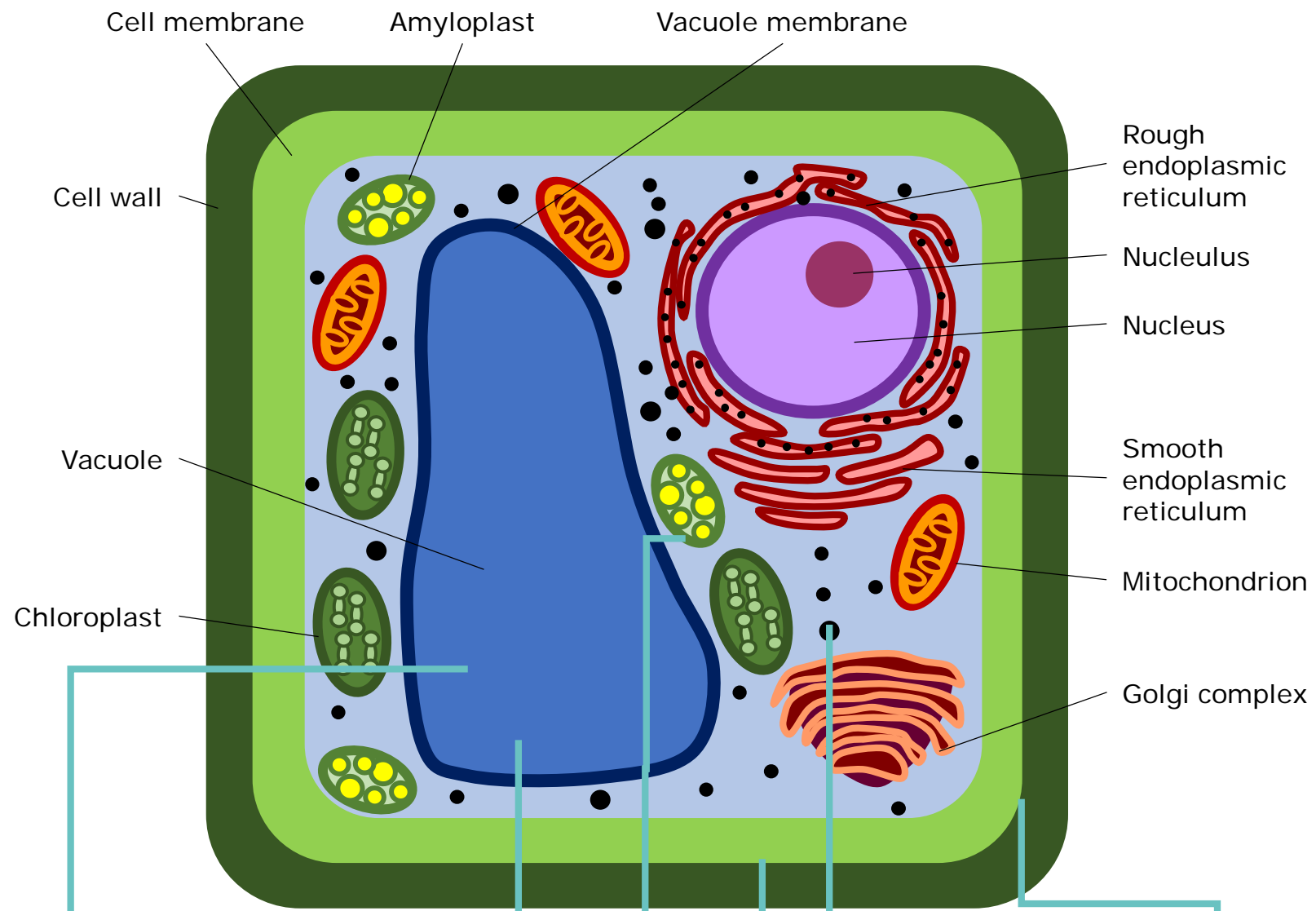
Wine production % (excluding juice and must)



OIV, International Organisation of Vine and Wine, 2020

Agri-food by-products: a rich and low-cost source of bioactive compounds





Cell membrane

Amyloplast

Vacuole membrane

Cell wall

Rough endoplasmic reticulum

Nucleolus

Nucleus

Vacuole

Smooth endoplasmic reticulum

Mitochondrion

Chloroplast

Golgi complex

Polyphenols

Proteins

Poly-saccharides

Lipids

Fibers



How to unlock and recover intracellular bioactives from these sources?

Need for cell disintegration pre-treatments

- Infusion
- Maceration
- Percolation
- Soxhlet extraction
- Decoction.



Strength:

- Well-established techniques
- Widely spread techniques.



Weaknesses:

- Energy intensive
- High solvent consumption
- Long extraction times
- High temperatures
- Low extraction yields.

• Pulsed electric fields (PEF)-assisted extraction

- High Pressure Homogenization (HPH)
- Ultrasound-assisted extraction (UAE)
- Supercritical fluid extraction (SFE)
- Microwave-assisted extraction (MAE)
- High Voltage Electrical Discharges (HVED)-assisted extraction.



Strength:

- Reduced residence times and solvent consumption
- Improved mass transfer
- Preserved functional properties
- Enhanced yields



Weaknesses:

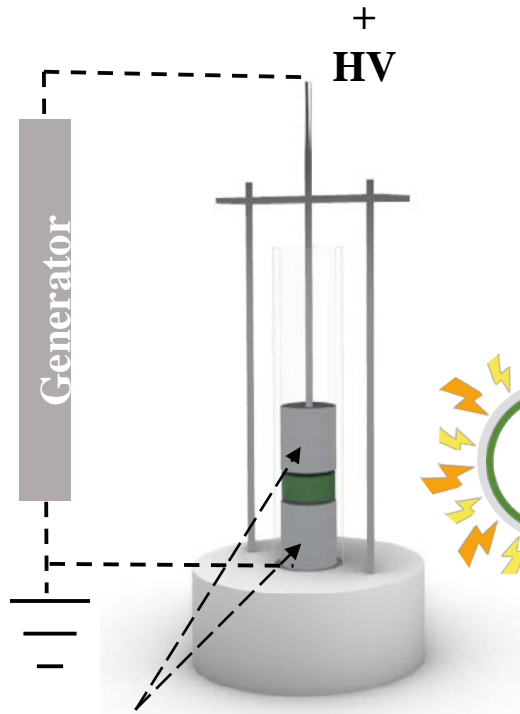
- Significant investment costs

Conventional extraction techniques

Non-conventional extraction techniques

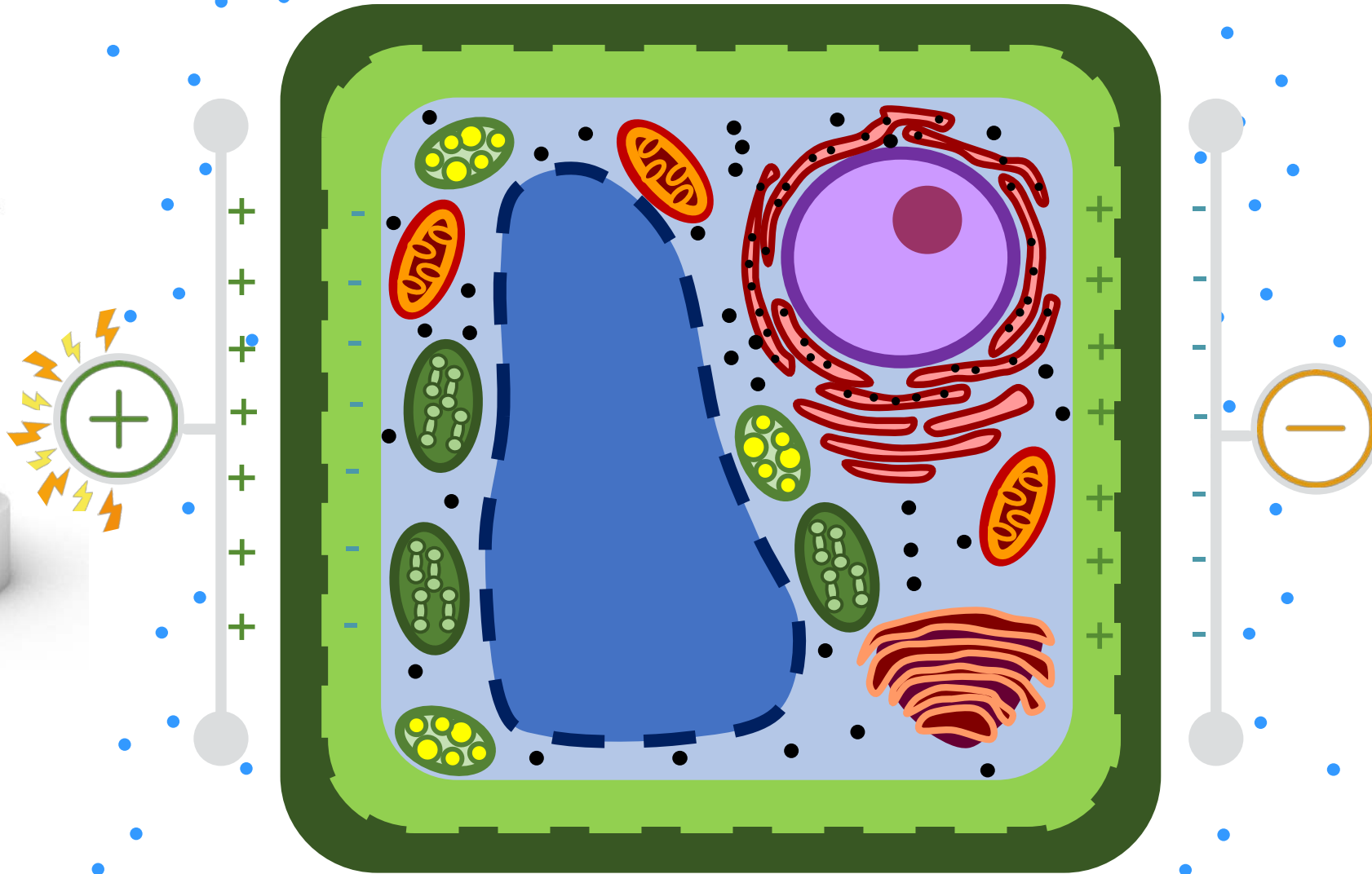


Pulsed Electric Fields: an effective cell permeabilization approach



Parallel Plate electrodes

E : 0.5–10 kV/cm
 W_T : 1–20 kJ/kg



Selective membrane permeabilization

Main challenges to face

Few works demonstrated the feasibility of PEF technology to intensify the recovery yield of phenolic compounds from winery by-products

None of them focused on the extractability of phenolic compounds from white grape pomace

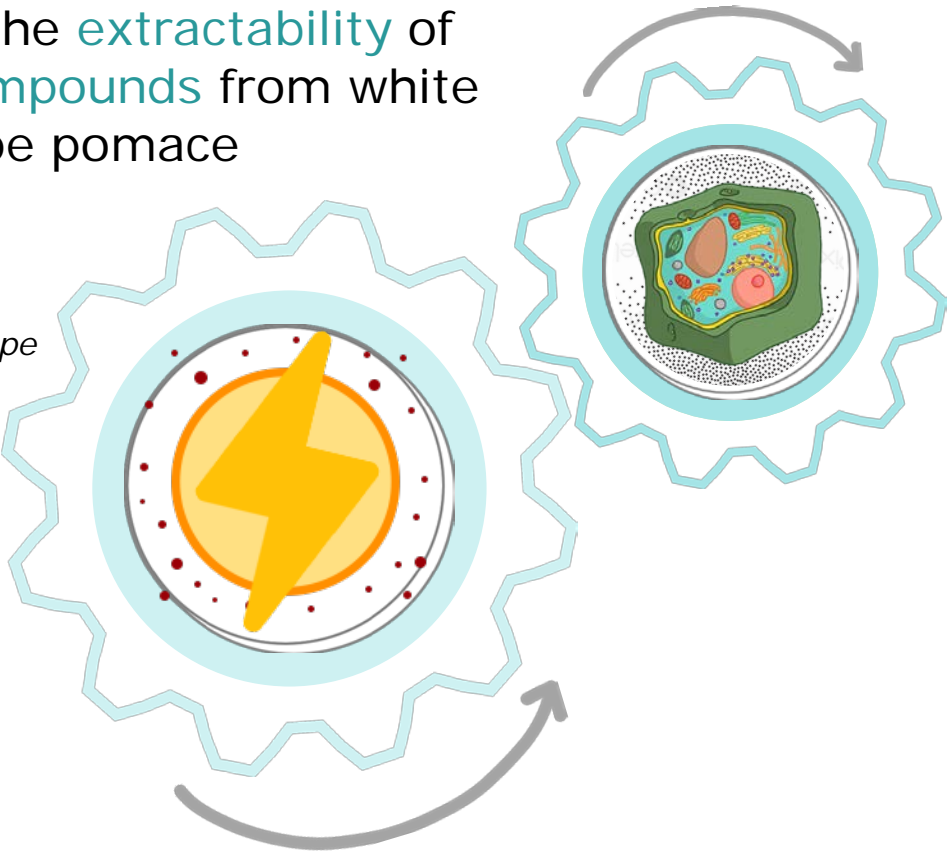
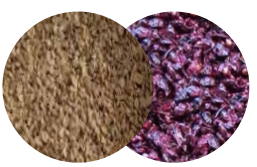
To fully exploit the advantages deriving from the application of PEF-assisted extraction over the conventional SLE process, an optimization step is required

No study on the optimization of the whole innovative extraction process made of a PEF pre-treatment stage followed by a subsequent SLE step

Objective of the work

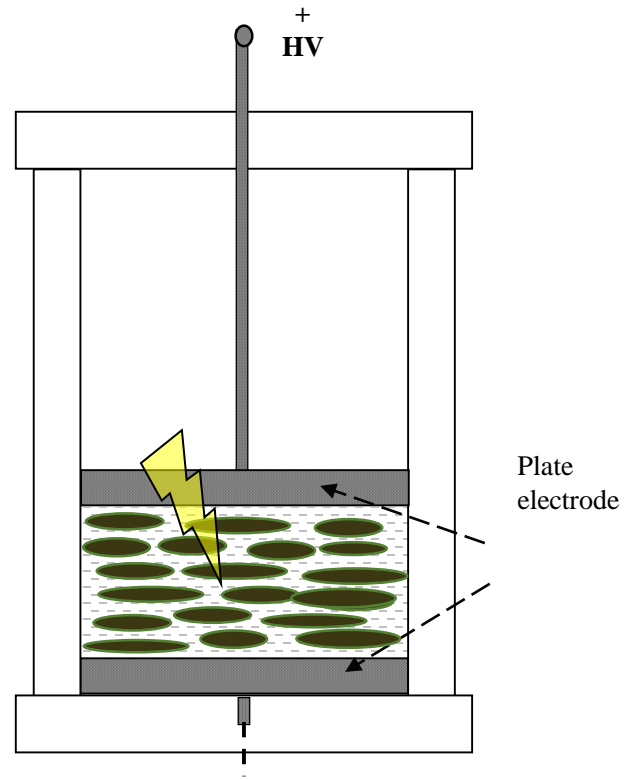
Potentiality of PEF pre-treatment to intensify the extractability of bioactive compounds from white and red grape pomace

White and Red grape pomace



Optimization of the PEF processing conditions based on cell disintegration index evaluations

PEF



Input variables

- $E=0.5-5$ kV/cm
- $W_T=1-20$ kJ/kg

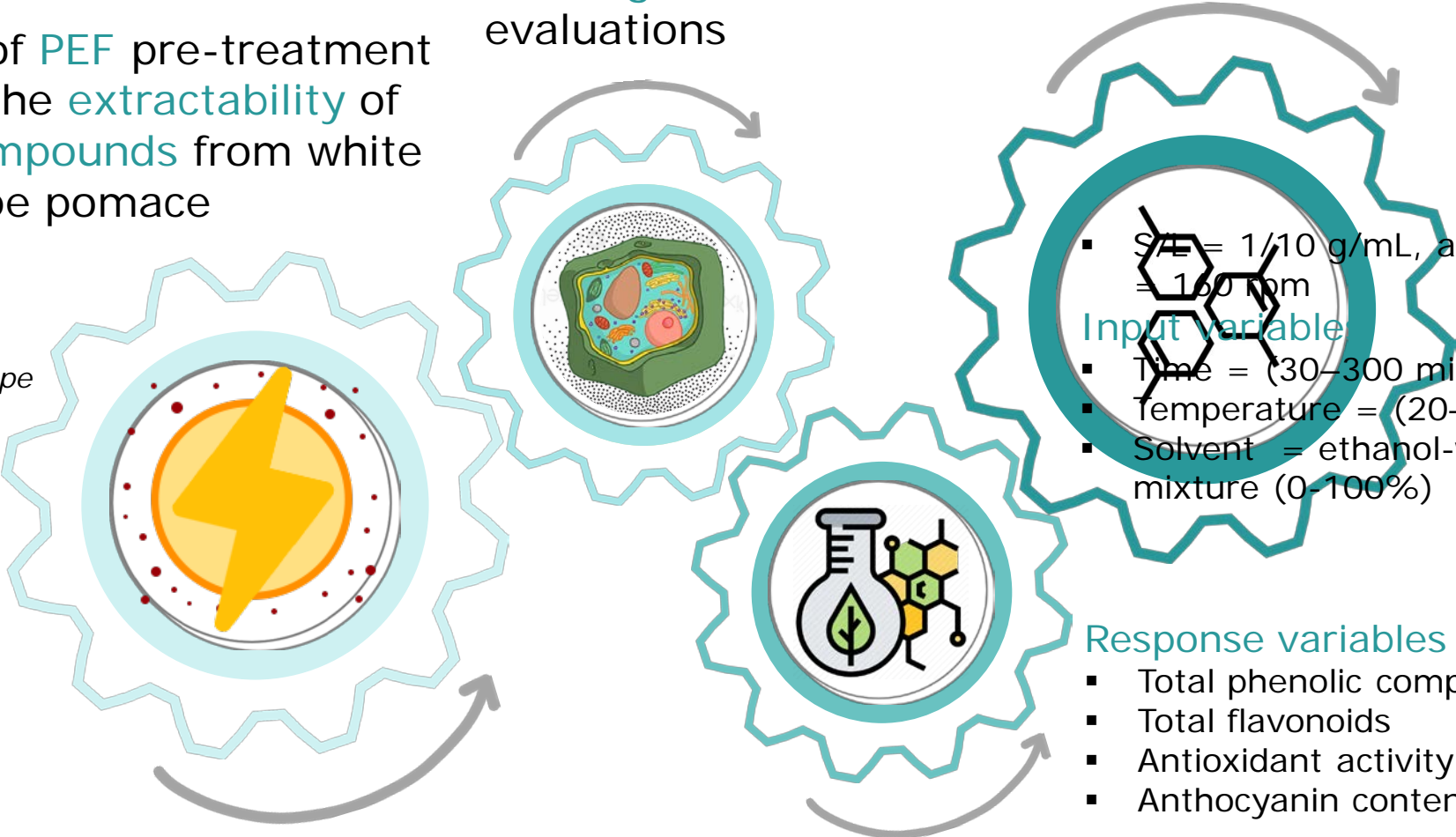
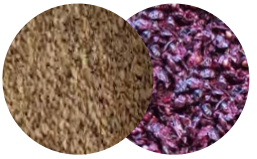
Response variable

- Z_p

Objective of the work

Potentiality of PEF pre-treatment to intensify the extractability of bioactive compounds from white and red grape pomace

White and Red grape pomace



Optimization of the PEF processing conditions based on cell disintegration index evaluations

Quantification of the Phenolic Compounds via HPLC-PDA Analysis

- Input variable
- $S/E = 1/10$ g/mL, agitation = 160 rpm
 - Time = (30–300 min),
 - Temperature = (20–50 °C),
 - Solvent = ethanol-water mixture (0–100%)

- Response variables
- Total phenolic compounds
 - Total flavonoids
 - Antioxidant activity
 - Anthocyanin content

Optimization of the SLE conditions



RESULTS

Optimization of PEF processing conditions

Cell Membrane Permeabilization of Grape Pomace Tissues

$$Y_k = \beta_0 + \sum_{i=1}^2 \beta_i X_i + \sum_{i=1}^2 \beta_{ii} X_i^2 + \sum_{i=1}^2 \sum_{j=i+1}^3 \beta_{ij} X_i X_j$$

X_i and X_j: independent variables; b₀, b_i, b_{ii}, and b_{ij}: intercept, regression coefficients of the linear, quadratic, and interaction terms of the model, respectively.

Run	Variables		Response		Z _p [-]	
	E [kV/cm]	W _T [kJ/kg]	White grape pomace	Red grape pomace	White grape pomace	Red grape pomace
1	0.5	1	0.083 ± 0.001a	0.033 ± 0.006a		
2	0.5	10.5	0.169 ± 0.003b	0.075 ± 0.002b		
3	0.5	20	0.281 ± 0.020c	0.13 ± 0.001c		
4	2.75	1	0.544 ± 0.011d	0.208 ± 0.003d		
5	2.75	10.5	0.681 ± 0.010e	0.512 ± 0.003e		
6	2.75	20	0.747 ± 0.010f	0.579 ± 0.003f		
7	5.0	1	0.758 ± 0.010f	0.417 ± 0.021e		
8	5.0	10.5	0.854 ± 0.006g	0.521 ± 0.007f		
9	5.0	20	0.807 ± 0.002g	0.70 ± 0.006h		
			β₀	-0.123806		-0.148433
			β₁ (E)	0.349245	***	0.240292
			β₂ (W_T)	0.017540	***	0.018985
			β₁₂ (E x W_T)	-0.001756	**	0.002172
			β₁₁ (E x E)	-0.035406	***	-0.028986
			β₂₂ (W_T x W_T)	-0.000229	ns	-0.000562
			p value of the model	< 0.0001	***	< 0.0001
			R²	0.9956		0.9545
			RMSE	0.0067		0.0071

The results are expressed as mean ± standard deviation (n = 2 for factorial and axial points; n = 3 for central point). Values with different lowercase letter within the same column are significantly different (p ≤ 0.05).

$$Z_p = \frac{|4 \ln tr(0.1 \text{ kHz})| - |4 \ln tr(0.1 \text{ kHz})|}{|Z_{10} - Z_{10}|}$$

ns, not significant for p > 0.05.
 significant for p ≤ 0.01; *significant for p ≤ 0.001.
 RMSE, Root Mean Square Error.

White grape pomace: E = 3,8 kV/cm; W_t = 10 kJ/kg

Red grape pomace: E = 4,6 kV/cm; W_t = 20 kJ/kg



$$Y_k = \alpha_0 + \sum_{i=1}^3 \alpha_i X_i + \sum_{i=1}^3 \alpha_{ii} X_i^2 + \sum_{i=1}^3 \sum_{j=i+1}^4 \alpha_{ij} X_i X_j + \sum_{i=1}^3 \sum_{j=i+2}^5 \alpha_{ij} X_i X_j + \sum_{i=1}^3 \alpha_{iii} X_i^3 + \sum_{i=1}^3 \sum_{j=i+1}^4 \alpha_{iij} X_i^2 X_j + \sum_{i=1}^3 \sum_{j=i+1}^4 \alpha_{ijj} X_i X_j^2$$

Y_k: predicted response variables; X_i and X_j: independent variables; α₀, α_i, α_{ii}, α_{ij}, α_{iij}, and α_{ijj}: intercept, regression coefficients of the linear, quadratic, and interaction terms of the model, respectively

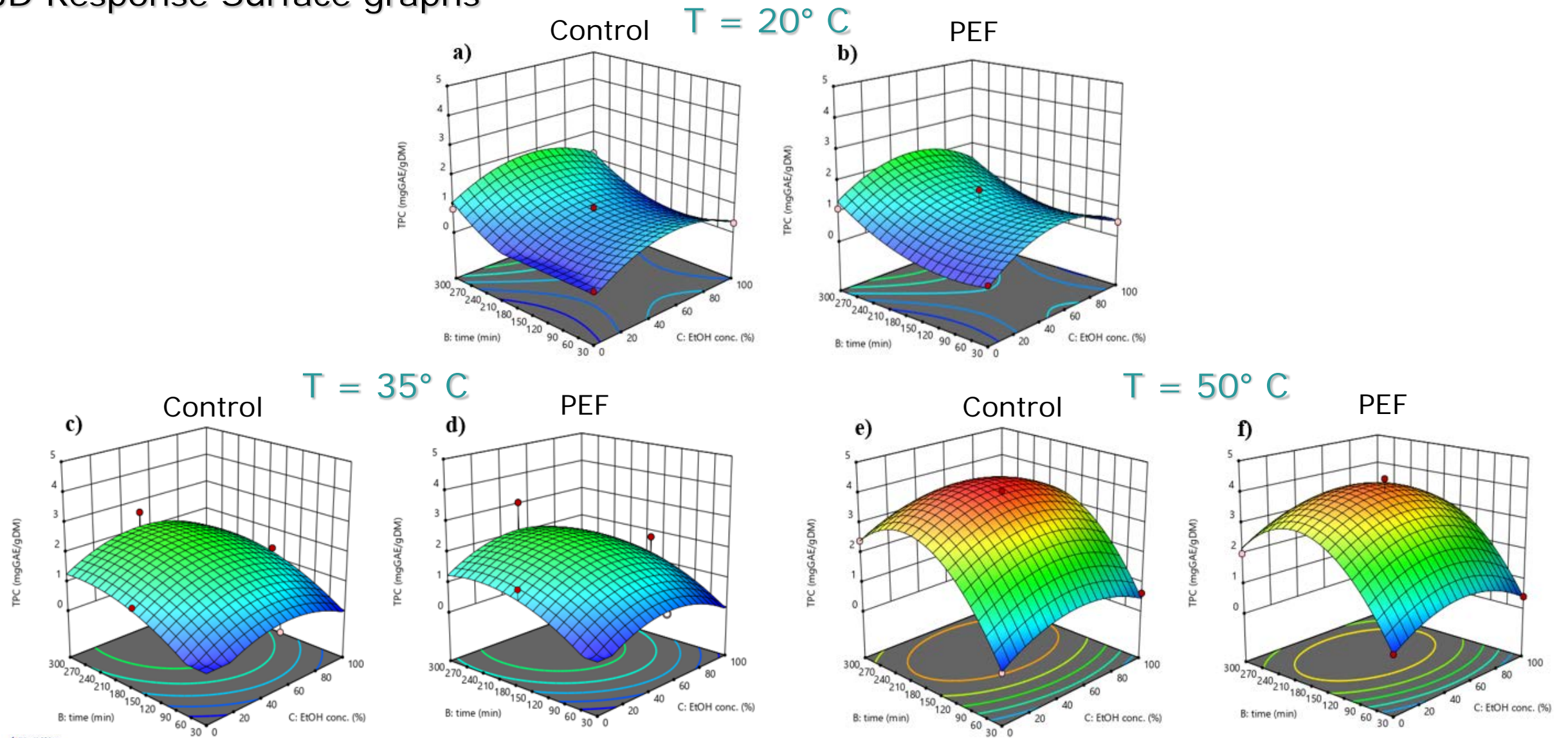
Run	SLE			PEF-assisted extraction		
	TPC (mgGAE/g _{DM})	FC (mgQE/g _{DM})	FRAP (mgAAE/g _{DM})	TPC (mgGAE/g _{DM})	FC (mgQE/g _{DM})	FRAP (mgAAE/g _{DM})
β₀	3.00885	5.60444	5.27926	3.3065	11.29163	6.57299
β₁ (T)	-0.187327	-0.380323 ***	-0.346407 ***	-0.198687 ***	-0.708839 ***	-0.414410 ***
β₂ (time)	-0.041053 ***	-0.054257 ***	-0.050758 ***	-0.036188 ***	-0.082916 ***	-0.056876 **
β₃ (EtOH)	0.038154 ***	0.022133 ***	0.027234 ***	0.04076 **	0.040509 **	0.036218 ns
β₁₂ (T x t)	0.001647 **	0.022157 **	0.003027 *	0.001539 ns	0.003215 ns	0.002992 ns
β₁₃ (T x EtOH)	-0.000041 ns	0.000332 ns	0.000214 **	-0.000094 ns	0.000371 ns	-0.000039 ns
β₂₃ (t x EtOH)	-0.000012 ns	0.000015 ns	-5.72614E-6 ns	-8.88358E-6 ns	0.000034 ns	-8.6925E-6 ns
β₁₁ (T x T)	0.002114 ***	0.004863 ***	0.004787 *	0.002297 **	0.008894 **	0.005638 ns
β₂₂ (t x t)	0.000129 **	0.000164 **	0.000059 **	0.000118 *	0.000253 *	0.000090 ns
β₃₃ (EtOH x EtOH)	-0.000321 *	-0.000202 *	-0.000301 ***	-0.000345 *	-0.000433 *	-0.000327 **
β₁₁₂ (T x T x t)	2.62 ± 0.07a	-2.54 ± 0.01a	-0.000034 2.74 ± 0.12a	3.06 ± 0.02b	3.00 ± 0.04b	0.000027 ± 0.10a
β₁₂₂ (T x t x t)	-4.55189E-6	-5.88399E-6	-2.15914E-6	-4.39068E-6	-9.22899E-6	-3.04141E-6
p value of the model	<0.0001 ***	<0.0001 ***	<0.0001 ***	0.0076 **	0.001249 ***	0.004254 **
R²	0.967	0.977	0.990	0.863	0.916	0.912
RMSE	0.057	1.193	0.095	0.345	0.651	0.153

TPC: +8%
FC: +31%
AA: +36%



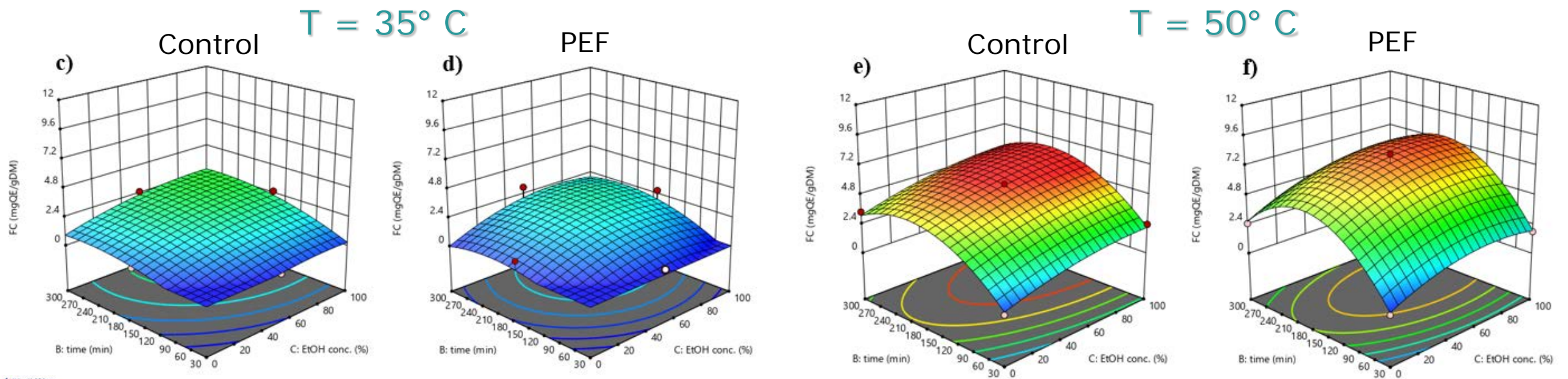
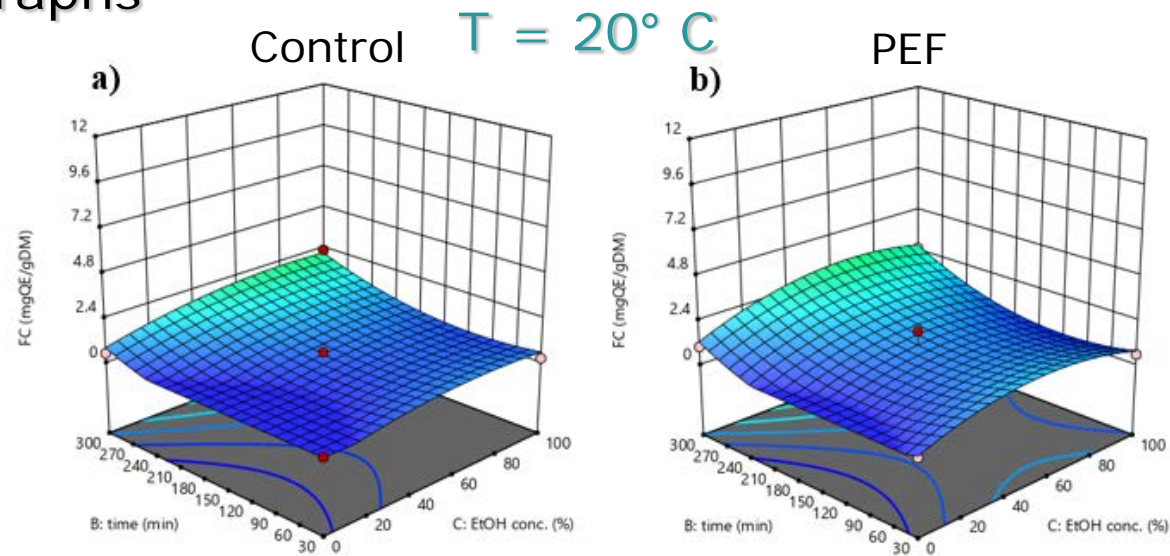
Total phenolic compounds (TPC)

3D Response Surface graphs



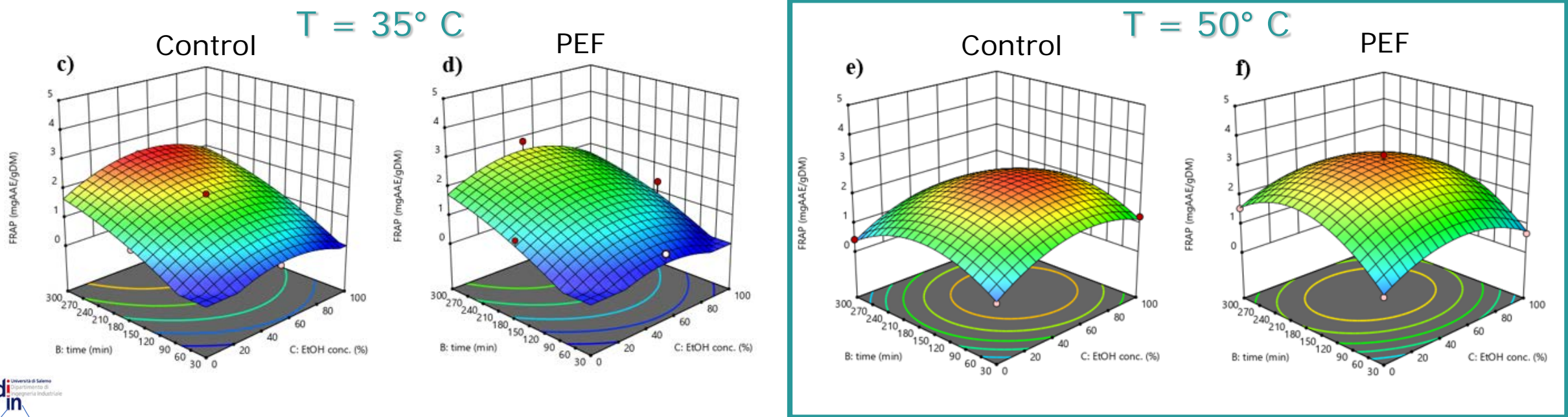
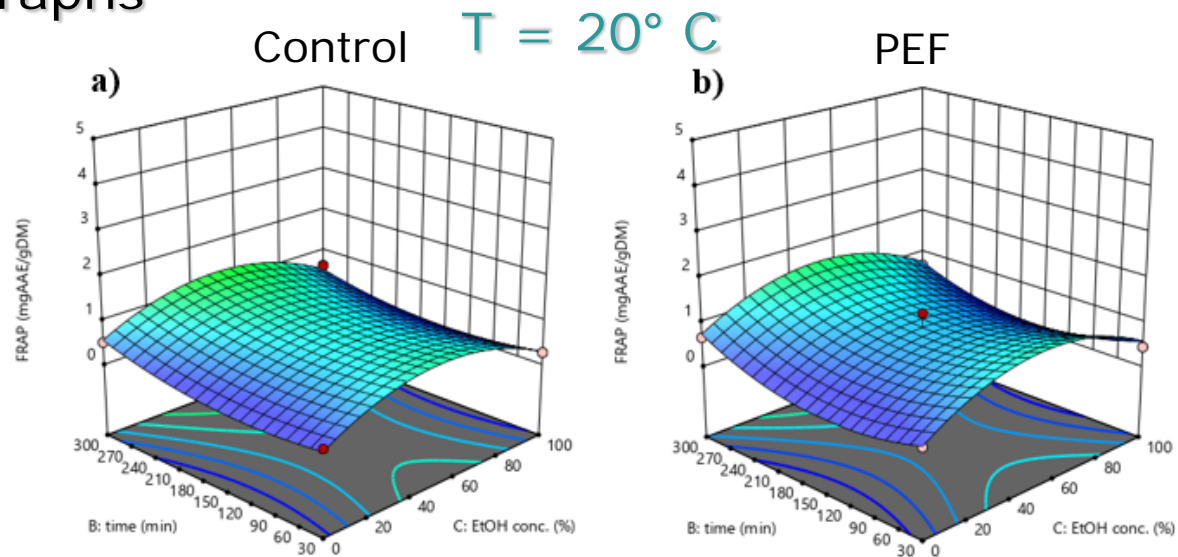
Flavonoid content (FC)

3D Response Surface graphs



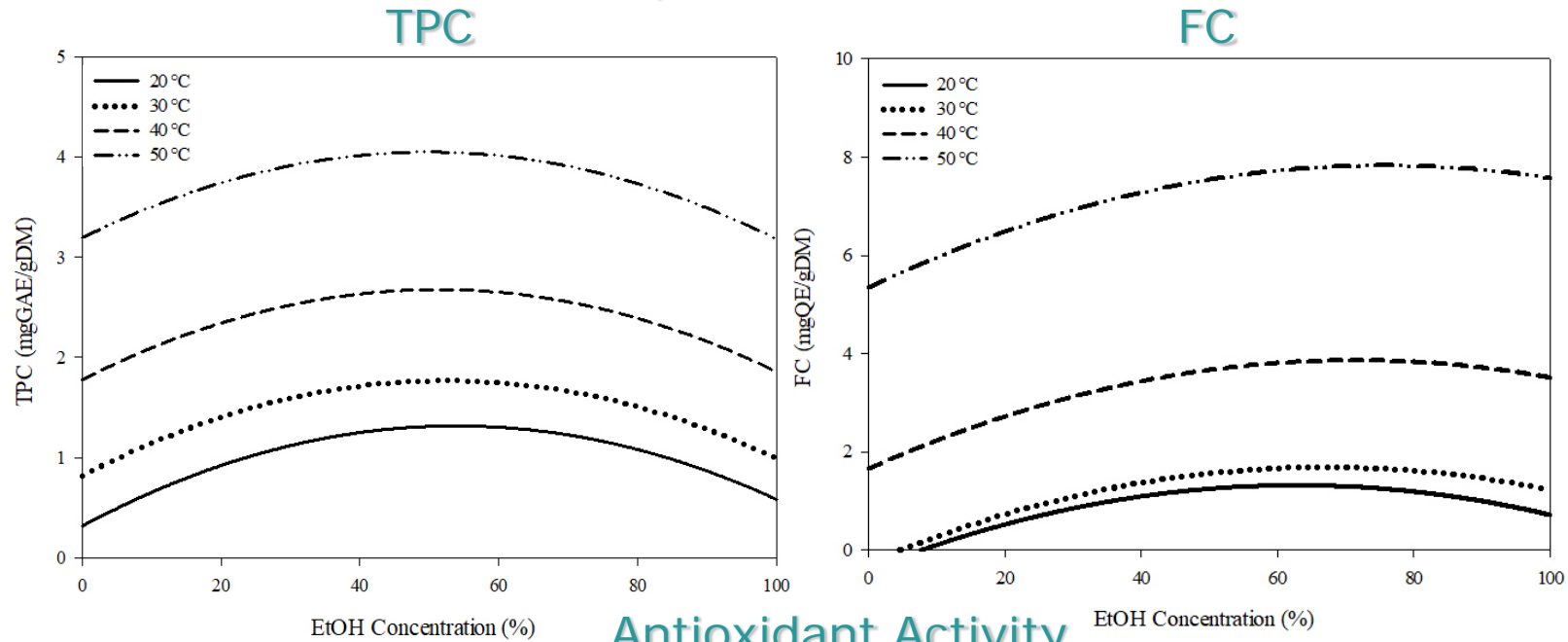
Antioxidant activity (FRAP)

3D Response Surface graphs

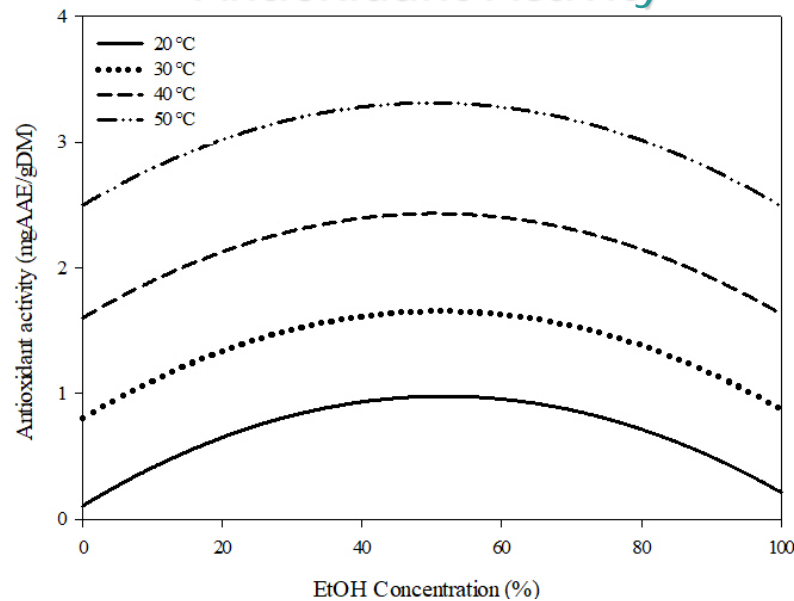


Pulsed Electric Fields (PEF): white and red grape pomace

Effect of the ethanol concentration on the response variables

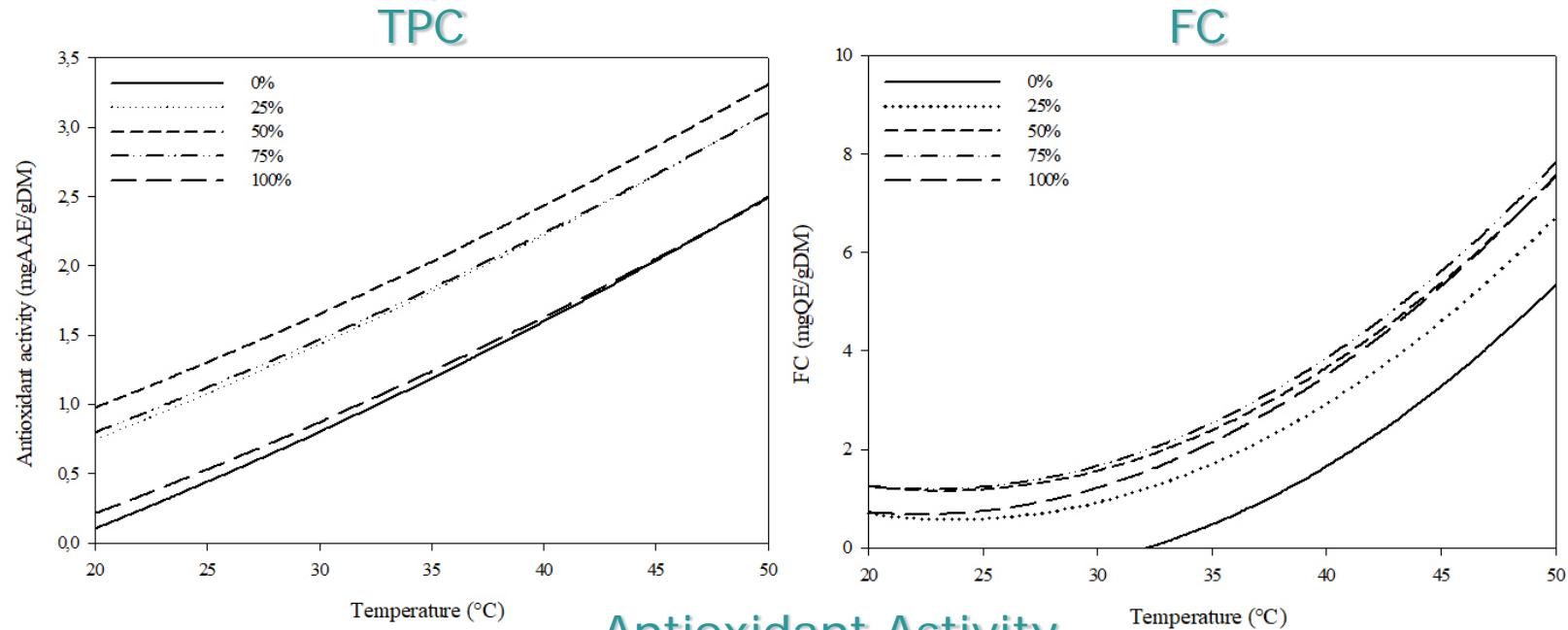


Antioxidant Activity

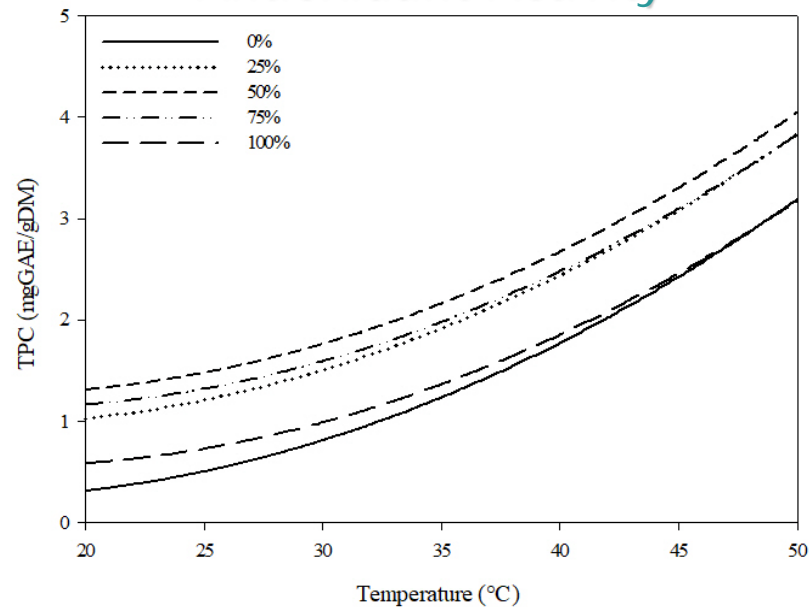


Pulsed Electric Fields (PEF): white and red grape pomace

Effect of the temperature on the response variables



Antioxidant Activity



Pulsed Electric Fields (PEF): red grape pomace

Response variables at the optimal PEF-assisted extraction conditions

Response	Temperature [°C]		Ethanol concentration [%]		Time [min]		Increment over SLE [%]			
	SLE	PEF	SLE	PEF	SLE	PEF	SLE	PEF		
TPC	50	50	53	50	300	213	190	+6%	+9%	
FC		50		50		300			+25%	
FRAP	50	50	88	76	223	192			+31%	
TAC	50	50	62	50	293	190			+54%	+36%

solvent consumption (3-12%)
extraction time (23-103 min)

Peleg's model fitting

$$TPC = \frac{t}{\frac{1}{v_0} + \frac{t}{TPC_\infty}}$$

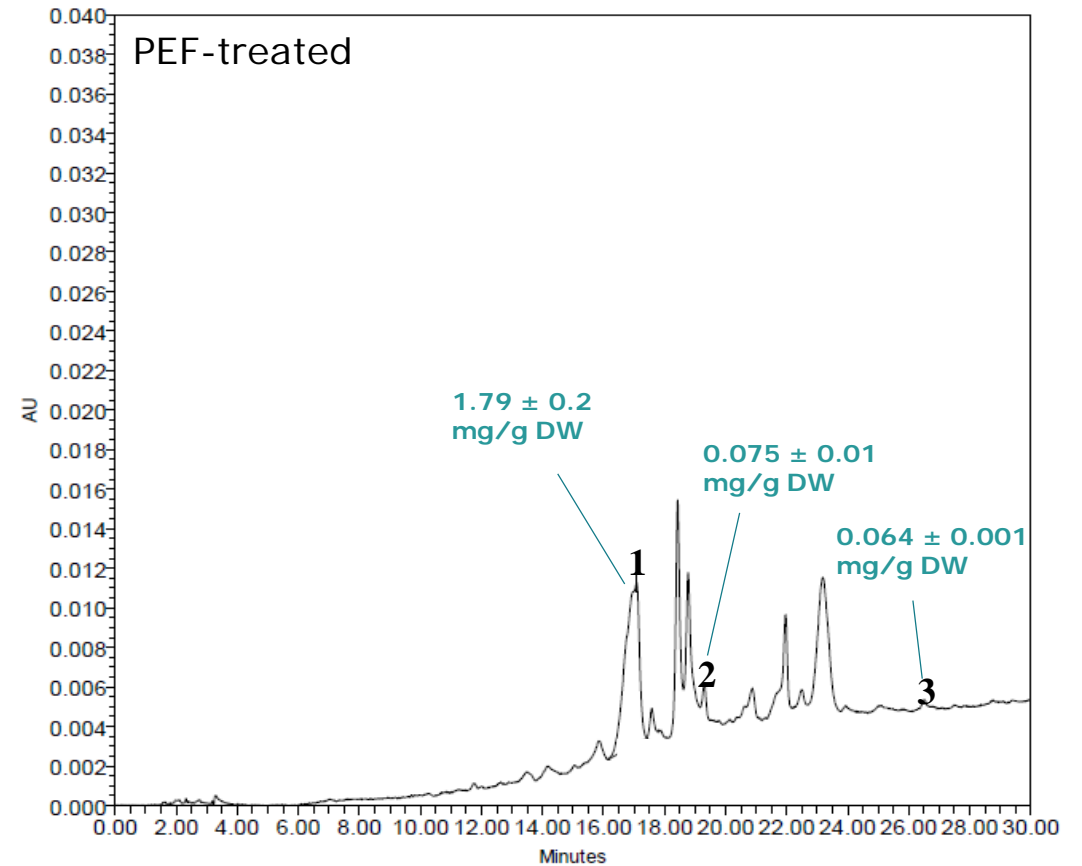
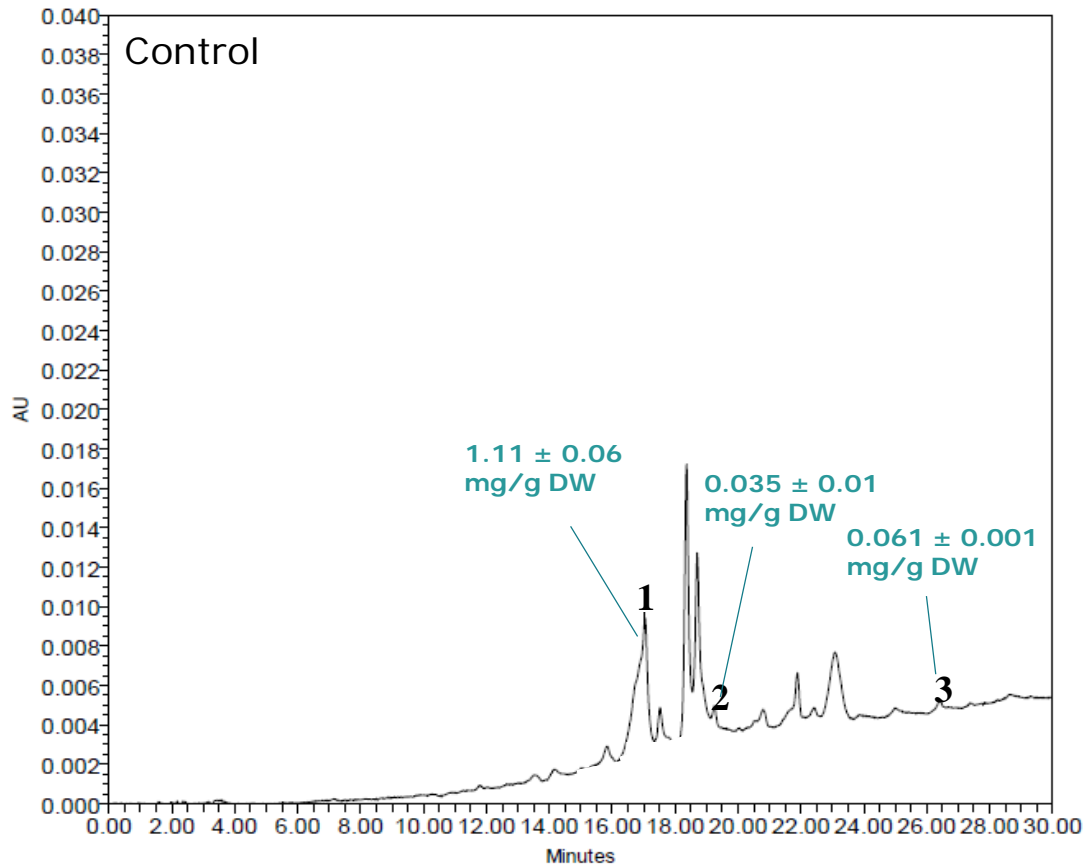
R² = 0.840-0.979

Selected overall optimal extraction conditions:
EtOH-Water = 50 % (v/v), T = 50 °C, Time = 300 min



Pulsed Electric Fields (PEF): white grape pomace

HPLC-PDA analysis: effect of PEF pre-treatment on the phenolic composition



Identified compound	Increment (%)
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Epicatechin	+61
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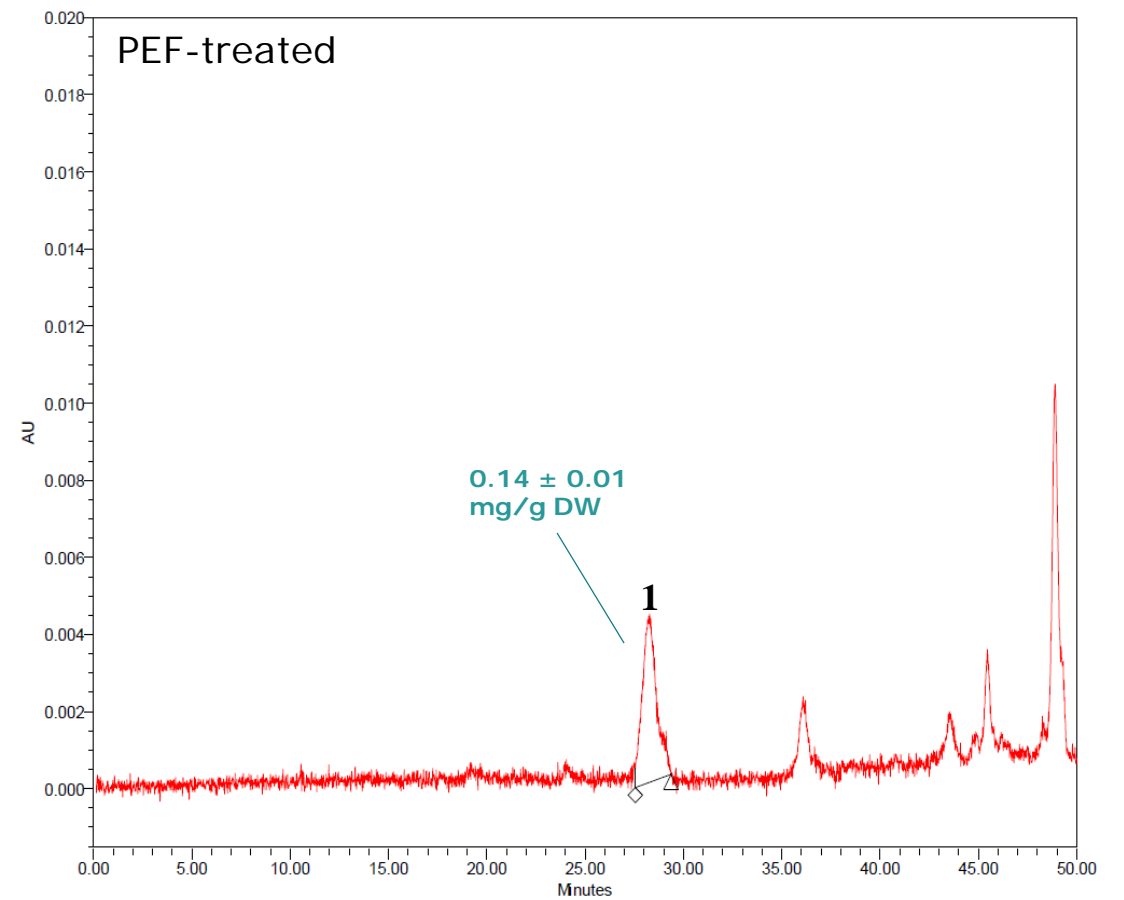
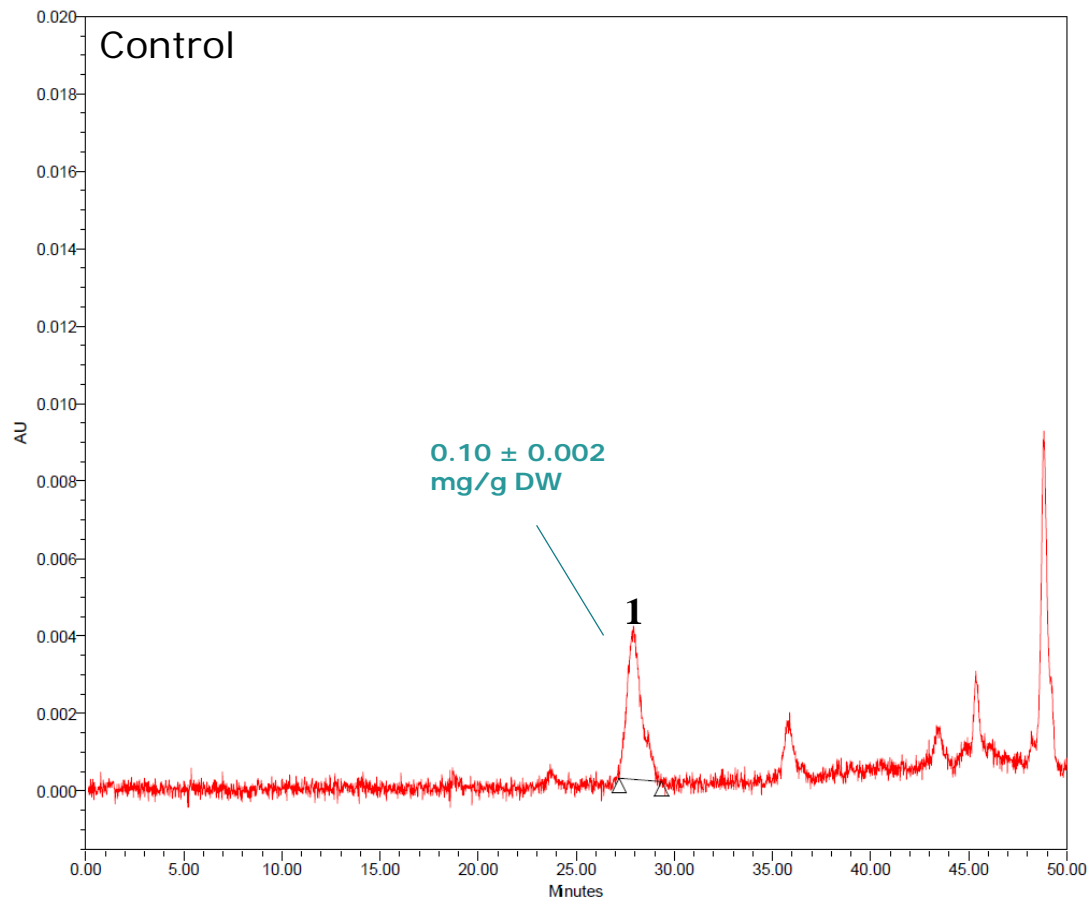
p-coumaric acid	+114
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Quercetin	+5
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1 = epicatechin; 2 = p-coumaric acid;
3 = quercetin

Pulsed Electric Fields (PEF): red grape pomace

HPLC-PDA analysis: effect of PEF pre-treatment on anthocyanin composition



Identified compound	Increment (%)
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Peonidin-3-O-glucoside

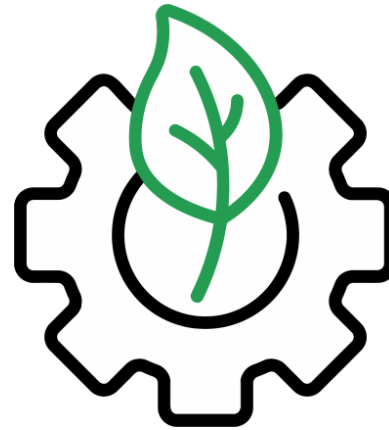
+40

1 = Peonidin-3-O-glucoside

Conclusions

Optimization step: the variables were significant, and the model accurately predicted the investigated responses for both PEF treatment and extraction step

Higher release of different phenolic compounds, including epicatechin (+61%), and p-coumaric acid (+114%) upon PEF compared to the control



PEF effective in intensifying the recovery yield of flavonoids (+24-31%), antioxidant power (+9-36%), and anthocyanin content (+40%) from grape pomace

Potentiality to reduce, the solvent consumption (3-12%) and shorten the extraction time (23-103 min) to achieve the same recovery yield of phenolic compounds



*Thank you
for your attention!*

