

Detoxification strategy of wheat straw hemicellulosic hydrolysate an approach for cultivating *Trichoderma reesei*

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(i) Introduction

र्दे Methodology

Detoxification stages Determination of toxicity thresholds

Cultivation of *T. reesei*

Results







Wheat production 2021: 778.6 Millions metric tons



Wheat straw Approx. 1.13 ton/ton grain









1. Detoxification stages
Finding conditions
Simulation software Aspen Plus V10

Evaporation synthetic hydrolysate (SH) / wheat straw hydrolysate (WSH)



Evaporation of WSH with pH modification

2. Determination of toxicity thresholds for *T. reesei*



Culture medium: Macro and micronutrients Glucose as carbon source (70 g/L)

+ Acetic acid Furfural HMF (individual analysis)



3. *T. reesei* cultivation in detoxified media





Fig 2. Monomeric and oligomeric sugars in wheat straw hydrolysate



Table 1. Detoxification of synthetic hydrolysate without pH modification

Compounds	Concentration [g/L]						
	Synthetic Hydrolysate	Concentrate	Condensate				
Glucose	0.24	7.40±0.39	N.D				
Xylose	0.85	26.64±1.43	N.D				
Acetic acid (AcH)	1.1	2.71±0.12	0.58 ±0.048				
Furfural	0.15	N.D	0.27 ± 0.022				
HMF	0.006	0.030±0.002	N.D				



 Table 2. Detoxification of wheat straw hydrolysate without pH modification

Common de		Concentration [g/L]		
Compounds	Wheat straw Hydrolysate	Concentrate	Condensate	
Total glucose	1.74	6.22 ± 0.47	N.D	
Total galactose	0.62	2.78 ± 0.13	N.D	55°C 140 mbar
Total mannose	0.27	1.95 ± 0.14	N.D	Initial volume evaporated: 70.6%
Total fructose	0.20	1.25 ±0.039	N.D	
Total xylose	8.97	31.80 ± 2.33	N.D	
Total arabinose	0.97	3.99 ± 0.29	N.D	
Acetic acid	1.40	3.50 ± 0.10	0.46 ± 0.11	
Furfural	0.41	N.D	0.44 ± 0.09	
HMF	0.021	0.086 ± 0.004.5	N.D	
 Similar trend t Fructose is deg Limited removies 	han synthetic hydrolysate graded val of acetic acid		Condensate: 100 % furfural 25% AcH	Concentration factors: Sugars 3.5 times 47.9 g/L total AcH 2.5 times HMF 4 times
		9th International Confe	rence on Sustainable Sol	id Waste

Management

Results detoxification. Stage 1: preliminary detoxifications using wheat straw hydrolysate (WSH)



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Fig 3. Acetic acid (AcH) and acetate (Ac-) equilibrium in a synthetic hydrolysate.

 Table 3. Detoxification of synthetic hydrolysate with alkaline and acid addition

Feed							
Feature	Units	SH + H ₂ SO ₄	SH + NaOH (1)	SH + NaOH (2)			
рН	-	2.00	4.66	5.46			
Total AcH	[mg/L]	1.46	1.46	1.46			
AcH	[mg/L]	1.44	0.81	0.24			
Ac⁻	[mg/L]	0.02	0.65	1.22			
% AcH	%	100	56	17			
	(Concentrate					
Total volume evaporated % 71.2 63.7 73.0							
Final pH	-	1.60	4.93	5.74			
Concentration total AcH	[mg/L]	2.79	3.54	5.06			
AcH	[mg/L]	2.78	1.43	0.48			
Ac⁻	[mg/L]	0.01	2.11	4.58			
Acetic acid removed	%	44.8	22.2	6.5			
Concentration factor AcH	times	1.9	2.4	3.5			
Concentration factor HMF	times	3.7	3.4	4.0			



Table 4. Detoxification of acidified wheat straw hydrolysate

C	С	Concentration [mg/L]			bution after ent [%]	
Compounds	Wheat straw Hydrolysate	Concentrate	Condensate	Concentrate	Condensate	
Total glucose	1.89	20.01 ± 7.1	N.D	100	0	55°C 140 mbar
Total galactose	0.91	9.83 ± 3.6	0.017± 0.010	98.2	1.8	Initial volume evaporated: 87.7 %
Total mannose	0.53	5.58 ± 1.88	N.D	100*	0	
Total fructose	0.11	0.68 ± 0.08	N.D	66.2	0	
Total xylose	11.72	117.1 ± 3.64	N.D	100	0	
Total arabinose	1.36	14.44 ± 4.80	N.D	100	0	
Acetic acid	1.74	7.45 ± 1.79	1.20 ± 0.014	44.1	55.9	Concentration factors
Furfural	0.41	0.012 ± 0.006	0.38 ± 0.08	0.3	99.7	Sugars 10 times
HMF	0.03	0.34 ± 0.14	N.D	100	0	
			9th International (Conference on S	Condensate 99.7 % furfu 55.9 % AcH Sustainable So	HMF 13 times
				Manageme	nt	8

Results. Determination of toxicity thresholds for *T. reesei*





Fig 4. Toxicity thresholds of a) furfural, b) acetic acid, and c) HMF.

- Furfural is the most harmful degradation product
- Acetic acid in low concentrations can be metabolized
- Detoxified wheat straw hydrolysate contain safe concentrations of furans.



3. *T. reesei* cultivation in detoxified media

Table 5. Bioma



Carbon sources (approx. 30 g/L): Glucose (control) Pre-treated SH and WSH

1. Detoxification stages



Biomass yield (g/g total sugars at the beginning)

HMF [[g/L]	Biomass			
Г _о	T _f	T _f [g]	Yield [g/g]*		
0	0	0.297	0.430		
026	0.01	0.014	0.017		
041	0	0.032	0.050		
045	0	0.045	0.078		
043	0	0.028	0.035		
036	0	0.230	0.420		

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Table 5. Biomass cultivation in different culture media

Culture media	Evap Vol. %	Initial sugars [g/L]	Sugars consumption %	Acetic acid [g/L]		HMF [g/L]		Biomass	
				Τ _ο	T _f	Τ _ο	T _f	T _f [g]	Yield [g/g]*
Control glucose	0	28	98.1%	0	0	0	0	0.297	0.430
SH	approx. 63-73	32	0.0%	2.4	2.0	0.026	0.01	0.014	0.017
SH + NaOH (2)		26	4.4%	2.6	0.1	0.041	0	0.032	0.050
SH + H₂SO₄		23	34.0%	2.4	1.7	0.045	0	0.045	0.078
WSH		32	2.1%	2.2	1.5	0.043	0	0.028	0.035
WSH + H ₂ SO ₄	approx. 88	22	60.3%	0.9	0	0.036	0	0.230	0.420

AcH effect (AcH and Ac⁻) over biomass production



•Wheat straw hydrolysate was detoxified, removing 99.7% furfural and 55.9% of acetic acid without sugar losses.

•Furfural in concentrations above 250 mg/L can cause severe inhibition in the cultivation of *T. reesei*.

•Acetic acid is produced in wheat straw hydrolysate at levels capable of impairing the production of *T. reesei* biomass.

•The acidification of wheat straw hydrolysis improved the removal of acetic acid and increased the production of biomass, reaching up to 98% of the yields in control.

Outlook...

Detoxified wheat straw hydrolysate can be used as a substrate for cultivating *T. reesei*. Further research is required towards the identification of possible products like enzymes.



S. Serna-Loaiza, F. Zikeli, J. Adamcyk, and A. Friedl, "Towards a wheat straw biorefinery: Combination of Organosolv and Liquid Hot Water for the improved production of sugars from hemicellulose and lignin hydrolysis," *Bioresour. Technol. Reports*, vol. 14, no. February, 2021, doi: 10.1016/j.biteb.2021.100667.

L. V. Daza Serna, C. E. Orrego Alzate, and C. A. Cardona Alzate, "Supercritical fluids as a green technology for the pretreatment of lignocellulosic biomass," *Bioresour. Technol.*, vol. 199, pp. 113–120, 2016, doi: 10.1016/j.biortech.2015.09.078.

K. Lu, N. Hao, X. Meng, Z. Luo, G. A. Tuskan, and A. J. Ragauskas, "Investigating the correlation of biomass recalcitrance with pyrolysis oil using poplar as the feedstock," *Bioresour. Technol.*, vol. 289, no. May, p. 121589, 2019, doi: 10.1016/j.biortech.2019.121589.

P. Phitsuwan, K. Sakka, and K. Ratanakhanokchai, "Improvement of lignocellulosic biomass in planta: A review of feedstocks, biomass recalcitrance, and strategic manipulation of ideal plants designed for ethanol production and processability," [15] Wang, J. Guo, P. Peng, M. Zhai, and D. She, "Hydrothermal degradation of hemicelluloses from triploid poplar in hot compressed water at 180-340 °c," *Polym. Degrad. Stab.*, vol. 126, pp. 179–187, 2016, doi: 10.1016/j.polymdegradstab.2016.02.003.

A. M. Borrero-López, E. Masson, A. Celzard, and V. Fierro, "Modelling the reactions of cellulose, hemicellulose and lignin submitted to hydrothermal treatment," Ind. Crops Prod., vol. 124, no. July, pp. 919–930, 2018, doi: 10.1016/j.indcrop.2018.08.045.

T. Dorđević and M. Antov, "The influence of hydrothermal extraction conditions on recovery and properties of hemicellulose from wheat chaff – A modeling approach," Biomass and Bioenergy, vol. 119, no. February, pp. 246–252, 2018, doi: 10.1016/j.biombioe.2018.09.030.

[G. Gallina, E. R. Alfageme, P. Biasi, and J. García-Serna, "Hydrothermal extraction of hemicellulose: from lab to pilot scale," Bioresour. Technol., vol. 247, no. July 2017, pp. 980–991, 2018, doi: 10.1016/j.biortech.2017.09.155.

D. Sun, Z. W. Lv, J. Rao, R. Tian, S. N. Sun, and F. Peng, "Effects of hydrothermal pretreatment on the dissolution and structural evolution of hemicelluloses and lignin: A review," *Carbohydr. Polym.*, vol. 281, no. August 2021, p. 119050, 2022, doi: 10.1016/j.carbpol.2021.119050.

S. Serna-Loaiza, M. Dias, L. Daza-Serna, C. C. C. R. de Carvalho, and A. Friedl, "Integral analysis of liquid-hot-water pretreatment of wheat straw: Evaluation of the production of sugars, degradation products, and lignin," Sustain., vol. 14, no. 1, 2022, doi: 10.3390/su14010362.

E. Jourdier, L. Poughon, C. Larroche, and F. Ben Chaabane, "Comprehensive study and modeling of acetic acid effect on trichoderma reesei growth," Ind. Biotechnol., vol. 9, no. 3, pp. 132–138, 2013, doi: 10.1089/ind.2013.0002.

M. Ivančić Šantek, M. Grubišić, M. Galić Perečinec, S. Beluhan, and B. Šantek, "Lipid production by Mortierella isabellina from pretreated corn cobs and effect of lignocellulose derived inhibitors on growth and lipid synthesis," *Process Biochem.*, vol. 109, no. May, pp. 46–58, 2021, doi: 10.1016/j.procbio.2021.06.021.

R. H. Bischof, J. Ramoni, and B. Seiboth, "Cellulases and beyond: The first 70 years of the enzyme producer Trichoderma reesei," Microb. Cell Fact., vol. 15, no. 1, pp. 1–13, 2016, doi: 10.1186/s12934-016-0507-6.

[M. Kolasa, B. K. Ahring, P. S. Lübeck, and M. Lübeck, "Co-cultivation of Trichoderma reesei RutC30 with three black Aspergillus strains facilitates efficient hydrolysis of pretreated wheat straw and shows promises for on-site enzyme production," *Bioresour. Technol.*, vol. 169, pp. 143–148, 2014, doi: 10.1016/j.biortech.2014.06.082.

[38] X. Liu *et al.*, "One-pot fermentation for erythritol production from distillers grains by the co-cultivation of Yarrowia lipolytica and Trichoderma reesei," *Bioresour. Technol.*, vol. 351, no. March, p. 127053, 2022, doi: 10.1016/j.biortech.2022.127053.

L. Shen, J. Gao, Y. Wang, X. Li, H. Liu, and Y. Zhong, "Engineering the endoplasmic reticulum secretory pathway in Trichoderma reesei for improved cellulase production," *Enzyme Microb. Technol.*, vol. 152, no. August 2021, p. 109923, 2022, doi: 10.1016/j.enzmictec.2021.109923.

C. Siamphan et al, "Production of D-galacturonic acid from pomelo peel using the crude enzyme from recombinant Trichoderma reesei expressing a heterologous exopolygalacturonase gene," J. Clean. Prod., vol. 331, no. January 2021, p. 129958, 2022, doi: 10.1016/j.jclepro.2021.129958.

M. Lindemann, A. Friedl, and E. Srebotnik, "Enhanced cellulose degradation of wheat straw during aqueous ethanol organosolv treatment," BioResources, vol. 12, no. 4, pp. 9407–9419, 2017, doi: 10.15376/biores.12.4.9407-9419.

B. Zhang et al., "High temperature xylitol production through simultaneous co-utilization of glucose and xylose by engineered Kluyveromyces marxianus," Biochem. Eng. J., vol. 165, no. September 2020, p. 107820, 2021, doi: 10.1016/j.bej.2020.107820.

S. Jo, J. Yoon, S. M. Lee, Y. Um, S. O. Han, and H. M. Woo, "Modular pathway engineering of Corynebacterium glutamicum to improve xylose utilization and succinate production," J. Biotechnol., vol. 258, pp. 69–78, 2017, doi: 10.1016/j.jbiotec.2017.01.015.

T. Ribeiro Correa, J. Barreto Roman, L. Vieira dos Santos, and G. Guimaraes Pereira, "Secretome analysis of Trichoderma reesei RUT-C30 and Penicillium oxalicum reveals their synergic potential to deconstruct sugarcane and energy cane biomasses," *Microbiol. Res.*, no. Ii, pp. 1–31, 2021, doi: 10.1016/j.micres.2022.127017.

National Center for biotechnology information, "PubChem Database," 2019. [Online]. Available: https://pubchem.ncbi.nlm.nih.gov/compound/Erythritol. [Accessed: 17-Jan-2020].

A. Cavka and L. J. Jornsson, "Comparison of the growth of filamentous fungi and yeasts in lignocellulose-derived media," Biocatal. Agric. Biotechnol., vol. 3, no. 4, pp. 197–204, 2014, doi: 10.1016/j.bcab.2014.04.003.







Thank you for you attention! Questions, comments?

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