

Quorum sensing system influences the synthesis of isoquercitrin catalyzed by recombinant *E. coli* BL21-pET21a-*rhaB1* biofilm

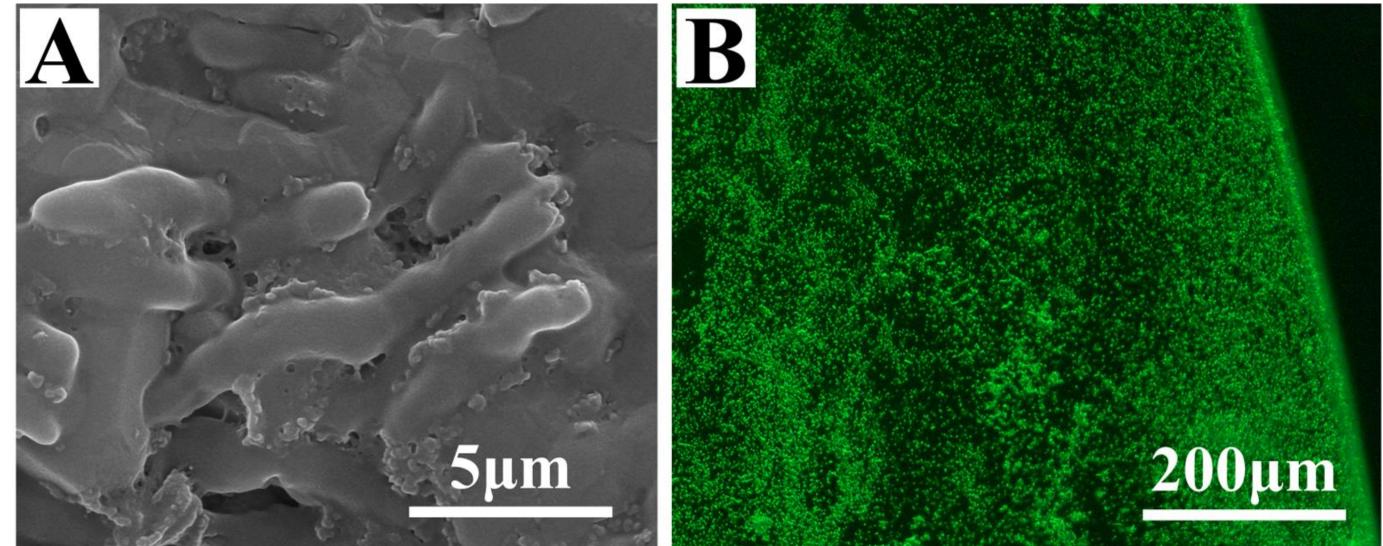
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Introduction

As flavonoids, isoquercitrin is an intermediate for the synthesis of new food additive Enzymatically Modified Isoquercitrin (EMIQ). But it is very rare in nature (1-5‰). However, isoquercitrin can be synthesized by hydrolyzing rhamnosyl of rutin, which can be found abundantly in nature. Biofilm is a structured community formed by the adhesion and aggregation of a large number of bacteria, and its advantages such as good self-regeneration, sustainability and scalability have great application potential in the field of biocatalysis. Previous studies have shown that quorum sensing system plays a key role in regulating biofilm formation. Therefore, it is of great significance to explore the relationship between quorum sensing and biofilm catalysis in engineering bacteria. Fig. 3 provides the observation of biofilm formation by scanning electron microscope (SEM) and fluorescence inversion microscope (FIM). recombinant *E. coli* biofilm was formed on a glass slide. Bacteria adhere to glass surfaces and secrete various highly organized, coordinated, and functional extracellular substances to form biofilms.





E. coli BL21-pET21a-*rhaB1* expressing RhaB1 were used to grow biofilm. Crystal violet was used to assess the total biomass assay of recombinant *E. coli* biofilm. *V. harveyi* BB170 was used as indicator bacteria to detect the activity of quorum sensing signal molecule AI-2. Finally, the biofilm was used to catalyze the synthesis of isoquercitrin from rutin.

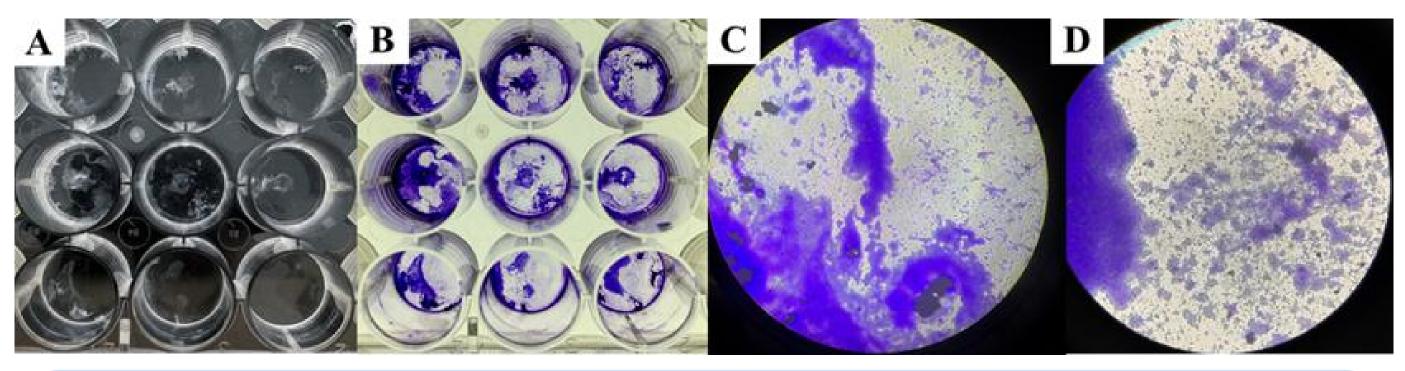


Fig. 1 Crystal violet staining and microscopic observation of biofilm. (A) Recombinant *E.coli* BL21-pET21a-*rhaB1* biofilm, (B) Crystal violet staining of the biofilm, (C) Microscopic observation of biofilm ($100 \times$), (D) Microscopic observation of biofilm ($400 \times$). Fig. 3 Observation of recombinant *E. coli* biofilm by SEM (A) and FIM (B).

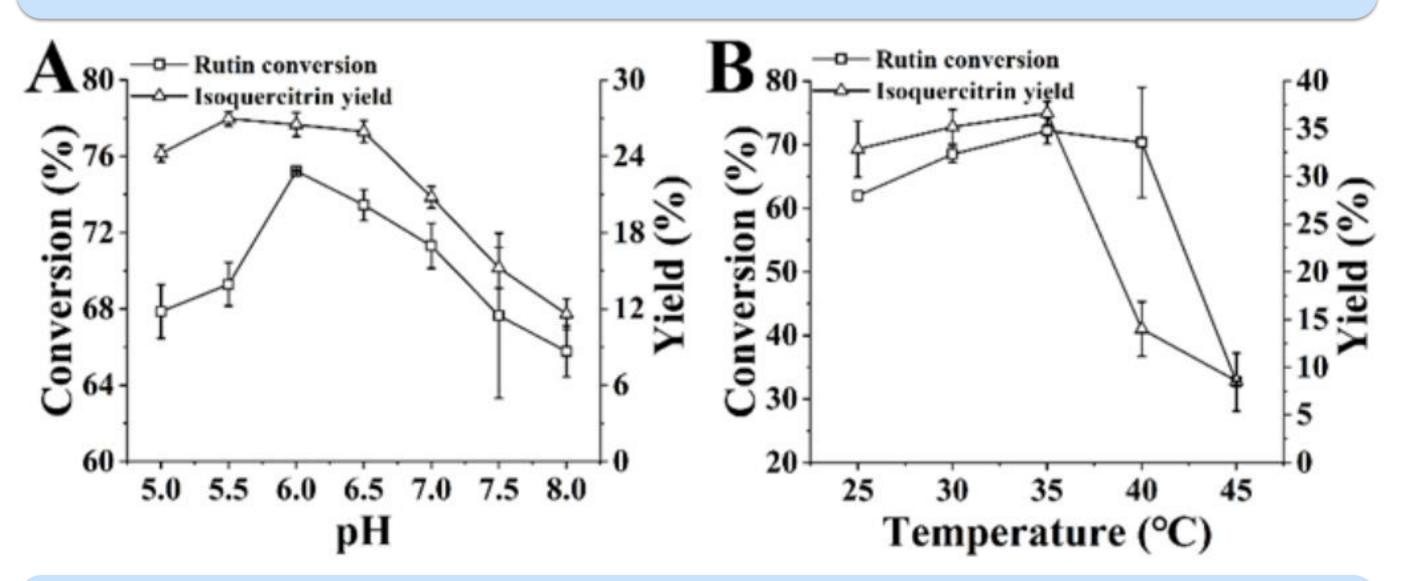


Fig. 4 Effect of pH (A) and temperature (B) on rutin conversion and isoquercitrin yield.

Results & Discussion

Fig. 2A shows that when pH increased from 3.5 to 8.5, the Al-2 activity of recombinant *E. coli* increased first and then decreased, and the activity was the highest at ph5.5. Fig. 2b monitored the dynamic changes of Al-2 activity in recombinant *E. coli* within 24 h. The Al-2 activity increased rapidly during the exponential growth period and decreased slowly after the stable period. Fig. 2C and Fig. 2D showed that the optimum growth condition of recombinant *E. coli* biofilm was pH 6.5 and temperature 35° C.

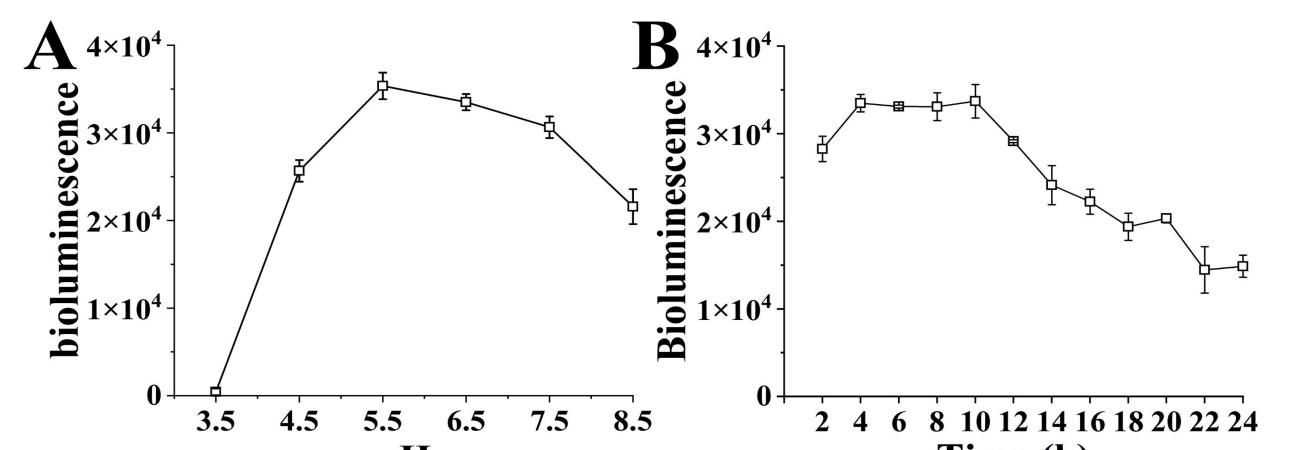


Fig. 4A shows that when pH is between 5.0 and 8.0, the rutin conversion rate increases first and then decreases, the maximum value is $75.22 \pm 0.05\%$, and the maximum isoquercitrin yield was $26.96 \pm 0.55\%$. Fig. 4B shows that the rutin conversion rate maintained a high level between 25°C and 40°C and then decreased rapidly, with the highest conversion rate reaching $72.20 \pm 8.67\%$, and the maximum isoquercitrin yield was $36.65 \pm 1.21\%$.

Conclusions

 AI-2 activity was highest at pH 5.5 and increased rapidly during the exponential growth period.

The optimum growth condition of recombinant *E. coli* biofilm is pH 6.5 and temperature 35°C.

• The acidic and low-temperature environment were more suitable for the reaction in the catalytic process of recombinant E. coli biofilm.

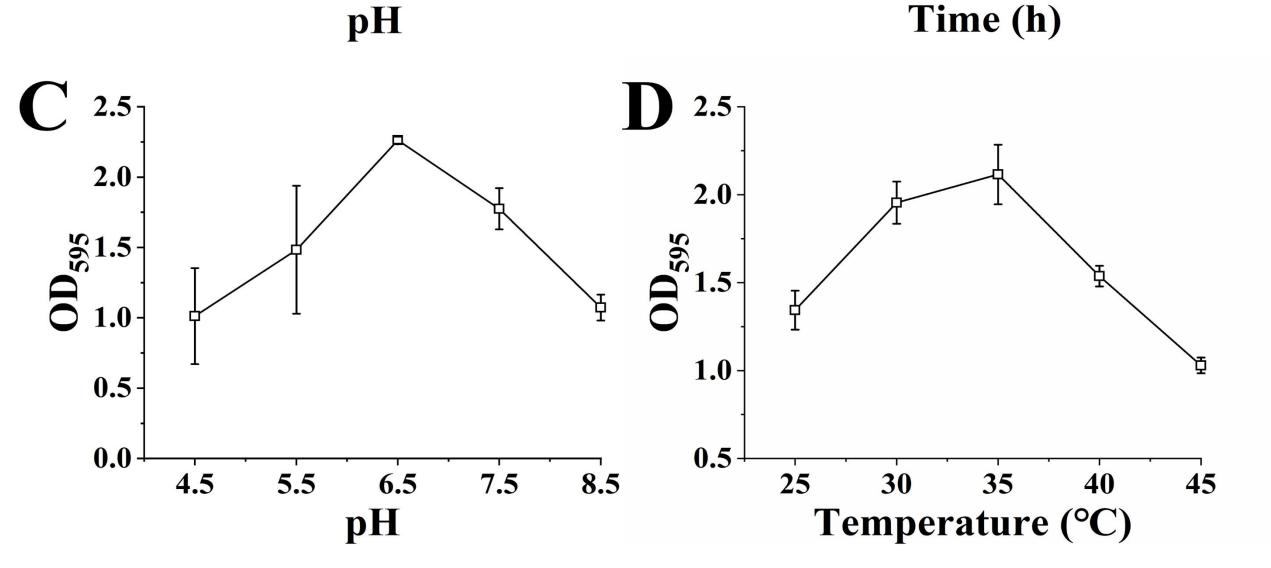


Fig. 2 Effects of environmental factors on QS system and biofilm biomass of recombinant *E. coli*. Effect of pH (A) and time (B) on AI-2 activity of QS system, Effect of pH (C) and temperature (D) on biofilm biomass.

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