## Tetracycline determines N<sub>2</sub>O fluxes through changes of microbial assemblages in a plant-soil system

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It is largely unknown how tetracycline results in changes to bacterial community composition and structure as well as in functional guilds involved in N-cycling when plants are present. In the current study, we examined the impact of tetracycline on N<sub>2</sub>O emissions, the abundance of bacterial guilds involved in N cycling in bare and planted with *Origanum officinalis* soil microcosms, and the bacterial composition in both bulk and root samples.

Overall, tetracycline had no effect on the abundance of the total bacterial community estimated as 16S rRNA copy numbers in the bulk soil. Tetracycline significantly affected the assembly of bacterial communities both in bulk and root samples. In bulk soil, differential abundance analysis showed that tetracycline caused preferential selection of specific taxa assigned in Firmicutes few days after its application. In bare soil, N<sub>2</sub>O fluxes were changing over time, which was associated with tetracycline concentration. Overall, low dose (0.1 mg/kg) application of tetracycline stimulated N<sub>2</sub>O emissions and led to an increase in the abundance of *nir*S and *nos*ZI bacterial communities.

On the contrary, in the plant-soil microcosms, we observed a substantial reduction in N<sub>2</sub>O daily emissions in soils treated with tetracycline, highlighting the importance of plant presence on N<sub>2</sub>O fluxes. The presence of antibiotic had no effect on the abundance of AOA or AOB. Redundancy analysis showed that the reduction of N<sub>2</sub>O fluxes was associated with a soil community harbouring nosZII genes while, the increase of daily N<sub>2</sub>O fluxes were mainly associated to the presence of NH<sub>4</sub><sup>+</sup>.