Growth characterization of adapted *Rhodosporidium toruloides* in sugarcane biomass hemicellulosic hydrolysate

Almeida, S.G.C., Souza, J.P., H.M. Fogarin, Franca, B.V., Dussán, K.J.

Department of Engineering, Physics and Mathematics, Institute of Chemistry, São Paulo State University-UNESP, Araraquara, São Paulo, Brazil

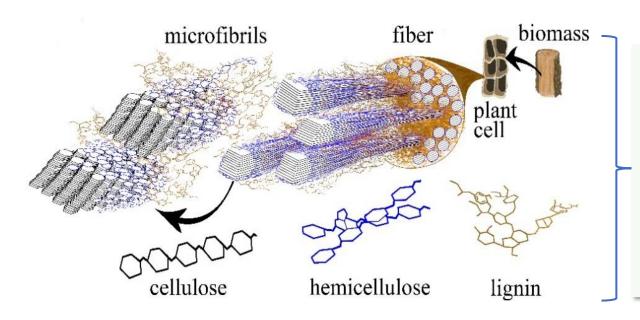


igement _____ Instituto de Químic Araraquar

10th International Conference on Sustainable Solid Waste Management 21-24 June 2023, Chania, Crete Island, Greece

Biomass

Scientific efforts have been demanded focus on to the diversification of the energy matrix, the obtainment of biomolecules and building blocks from renewable sources.



The sugars, present in the cellulosic and hemicellulosic portions of the biomass, represent more than **60%** of the biomass composition and the others **40%** being composed mainly of lignin, a polyphenolic biomolecule.

de Almeida, Sâmilla GC et al. 2022. Chapter: Methods for hemicellulose deconstruction aiming to xylose recovery: Recent progress and future perspectives

Biomass

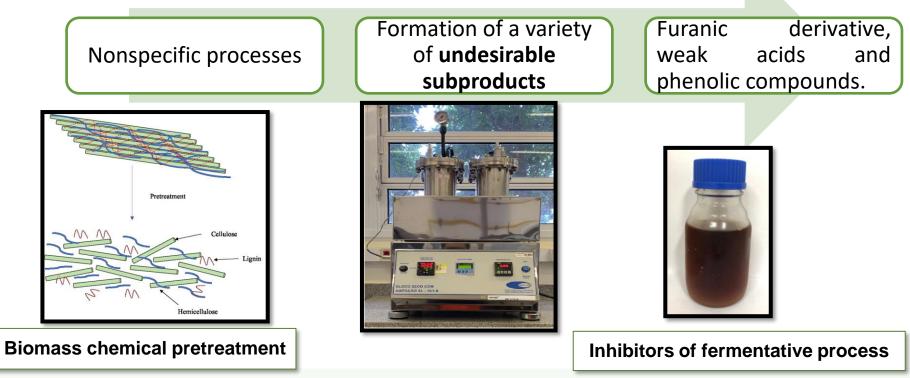


Valorization of agro-industrial waste, and lignocellulosic biomass has emerged as a potential raw material because of its availability, and capacity to be processed to obtain value-added products.

Biofuels

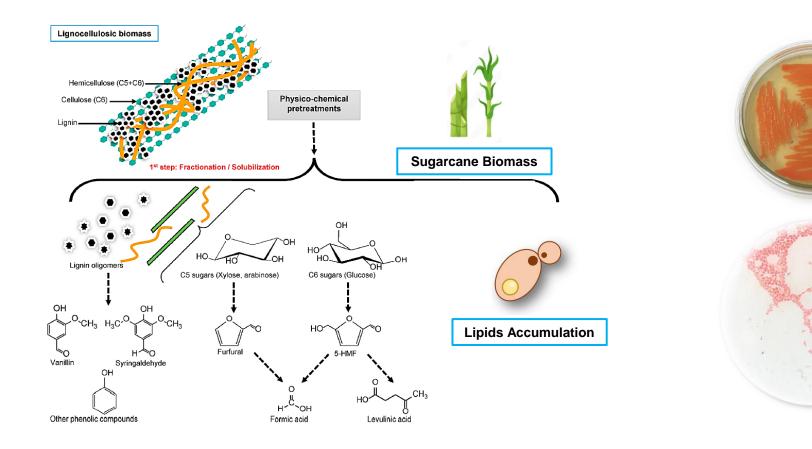
Functional Ingredients

Building blocks



Adaptation: alternative to replace the detoxification process and consists in the microorganism acclimation into gradually increased concentrations of the hydrolyzed in the presence of the toxic compounds, inducing the microorganism to adapt itself in the growth medium through its innate capability, which is the result of a selective pressure by the medium and the development of a metabolic apparatus.

- Bioprocess
- □ The oleaginous yeasts can consume the xylose from a wide range of materials ⇒ they are able to grow in the presence of compounds that breaks down the biomass, being able to metabolize these compounds present in the hemicellulosic hydrolysate ⇒ favoring the implementation of these types of bioprocesses.





Goals



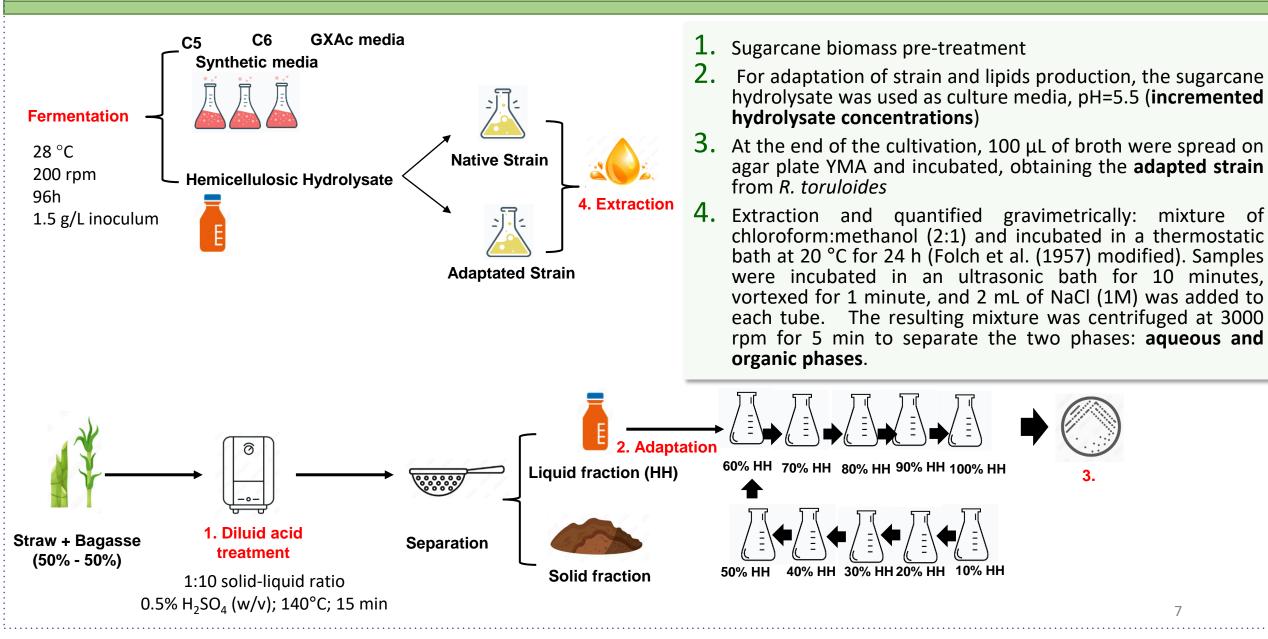
Exploitation of sugarcane straw and bagasse to produce lipid via yeast by exploring the:

- ✓ Three strains were cultivated in synthetic media to assess the pattern of consumption of carbon sources and strain growth
- ✓ Adaptative strategy in hemicellulosic hydrolysate
- The growth profile, lipid production of adapted and no-adapted cultivated in the non-detoxified hemicellulosic hydrolysate were compared for microbial lipid production

Approach

* **Fermentation**

mixture of



 Evaluation of the potential of R. toruloides to utilize C5 and C6 as carbon sources on synthetic media

Table 1. Comparisor	of lipid production	by R. toruloides o	n synthetic media
---------------------	---------------------	--------------------	-------------------

Yeast	рН _і	рН _f	Biomass (g.L ⁻¹)	Consumption of C (%)	Lipid accumulation % (m.m ⁻¹)	Yield biomass on carbohydrate Y _{X/S} (g.g ⁻¹)	Yield lipid on carbohydrate Y _{P/S} (g g ⁻¹)	Lipid production rate QP (mg. L ⁻¹ .h ⁻¹)
C5								
2781	4.68	3.03	7.09	30.74	3.92%	0.366	0.0168	2.89
2882	5.03	3.54	7.30	26.54	4.87%±0.770%	0.428	0.0248	3.706±0.5840
2896	5.16	4.46	11.44	41.26	4.69%±0.350%	0.492	0.0260	5.595±0.4169
C 6								
2781	4.99	2.57	10.86	34.91	6.16%±0.590%	0.493	0.000294	0.059±0.0057
2882	5.27	3.03	10.92	32.81	7.42%±0.506%	0.571	0.000396	0.071±0.0048
2896	5.22	2.69	9.74	36.69	6.70%±0.177%	0.432	0.000359	0.072±0.0019

□ The three strains of *R. toruloides* were cultivated in synthetic media to compare the ability of the strains to metabolize different sources of sugars

- □ The only strain that showed greater growth capacity in xylose (C5) was *R. toruloides* 2896
- □ *R. toruloides* 2781 and *R. toruloides* 2882 showed more expressive growth in glucose (C6)

 Evaluation of the potential of R. toruloides to utilize C5 and C6 as carbon sources on synthetic media

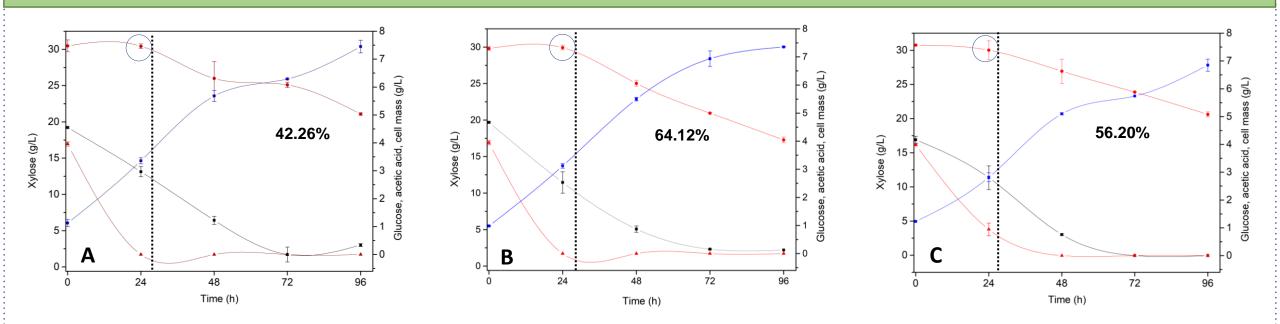


Fig. 1. Fermentation of synthetic medium with glucose, xylose and acetic acid (MGXAc) by (A) *R. toruloides* 2882, (B) *R. toruloides* 2896 and (C) *R. toruloides* 2781 at 28°C and 200 rpm. - ▲ - Glucose, -●-Xylose, -■- acetic acid, -■- cell mass

- In synthetic medium containing acetic acid (GXAc media): simultaneous consumption of glucose and xylose was not observed.
- □ Xylose consumption only started after 24 hours, with the exhaustion of glucose.

Adaptation yeast: increasing resistance to growth

 \mathbf{X}

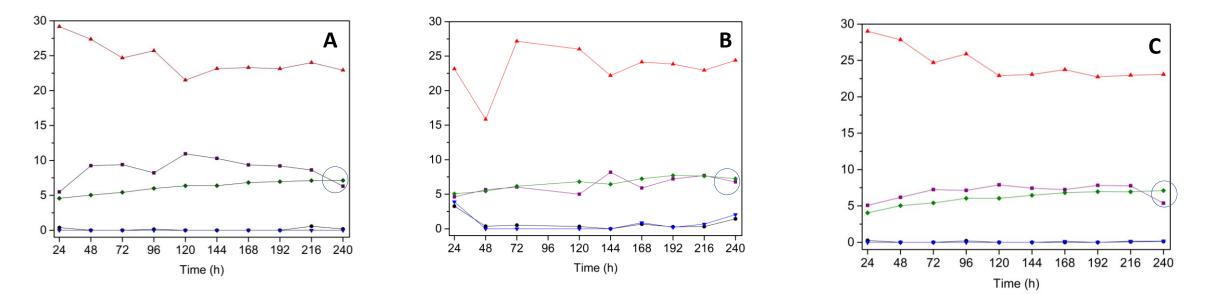
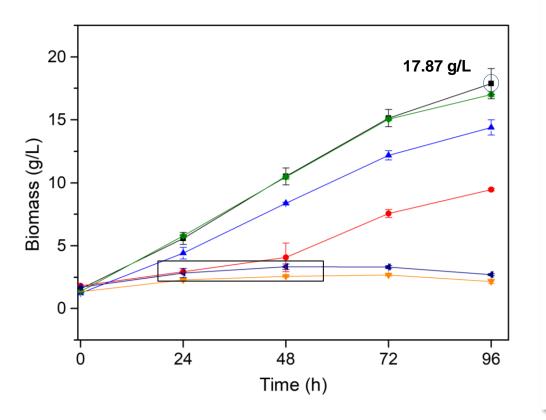


Fig. 2. Temporal profile of biomass accumulation (-■-), pH (-♦-), consumption of glucose (-●-), xylose (-▲-) and acetic acid (-▼-) by *R. toruloides* 2882 (A), *R. toruloides* 2896 (B) and *R. toruloides* 2781 (C) during the adaptation phase in HHB

- □ All strains were able to grow in 100% supplemented hydrolysate
- The cell concentration reached at the 10th level of adaptation (100% HHB) was higher to *R. toruloides* 2896, followed by *R. toruloides* 2882 and *R. toruloides* 2781
- In each adaptation cycle (lasting 24 hour) glucose was depleted as a preferred carbon source, for all strains

Comparison between R. toruloides adapted and non-adapted in HHS



Fermentative performance of non-adapted and adapted strains of *Rhodosporidium toruloides* in non-detoxified HHS supplemented:

- non-adapted strains: showed a lag phase up to at least 48 hours
- adapted strains: showed an exponential growth phase

**

R. toruloides 2882: presented exponential phase, with higher accumulation of cell biomass in both cases (adapted and non-adapted)

Fig. 3. Cell growth of *R. toruloides* in HHS non-detoxified supplemented non-adapted cells: -●- *R. toruloides* 2882, - ▼ - *R. toruloides* 2781, - ◀ - *R. toruloides* 2896 adapted cells: -■- *R. toruloides* 2882, - ▲ - *R. toruloides* 2781, -♦- *R. toruloides* 2896

Comparison between R. toruloides adapted and no adapted in HHS

*

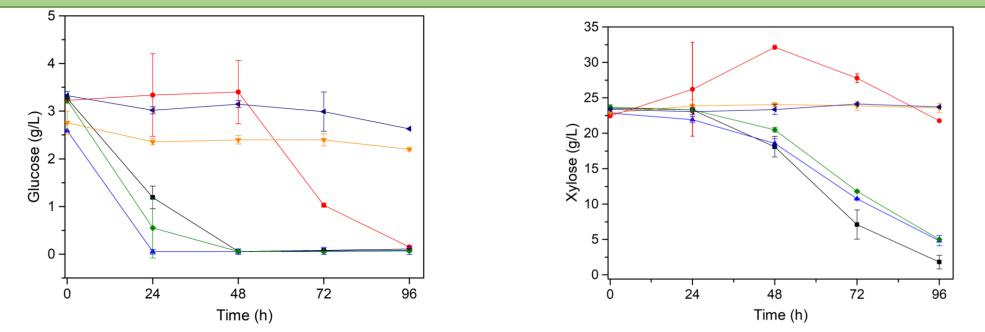


Fig. 4. Glucose and xylose consumption by *R. toruloides* grown in HHS non-detoxified supplemented non-adapted cells: -●- *R. toruloides* 2882, -▼- *R. toruloides* 2781, - **4**- *R. toruloides* 2896 Adapted cells: -■- *R. toruloides* 2882 - ▲ - *R. toruloides* 2781 - ◆- *R. toruloides* 2896

Consumption of sugars was improved in the adapted cells: glucose consumption occurred in the first 24 hours;
xylose consumption occurred after glucose depletion, observing its exhaustion in 96 hours of cultivation

The consumption of sugars (glucose and xylose) by non-adapted cells was 10%, 24% e 8.18% for strains 2781, 2882 e 2896, respectively. For adapted cells, the consumption of total sugars was 83%, 93.31% e 82% for strains 2781, 2882 and 2896

*

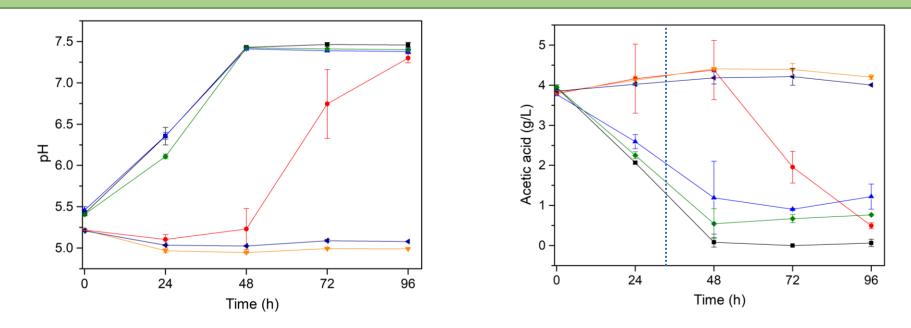
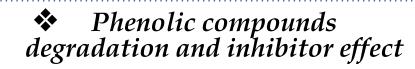


Fig. 5. Acetic acid consumption and pH variation during growth of *R. toruloides* in HHS non-detoxified supplemented non-adapted cells: -●- *R. toruloides* 2882, - ▼ - *R. toruloides* 2781, - ◀ - *R. toruloides* 2896 adapted cells: -■- *R. toruloides* 2882, - ▲ - *R. toruloides* 2781, -♦- *R. toruloides* 2896

□ With the exception of the non-adapted strains 2781 and 2896, the ability of *R. toruloides* to assimilate the acetic acid presented in the hydrolysate (as a carbon source) was observed. The decrease in acid concentration promoted the neutralization of broth, pH was increased from **5.5** up to a value of **7.5**



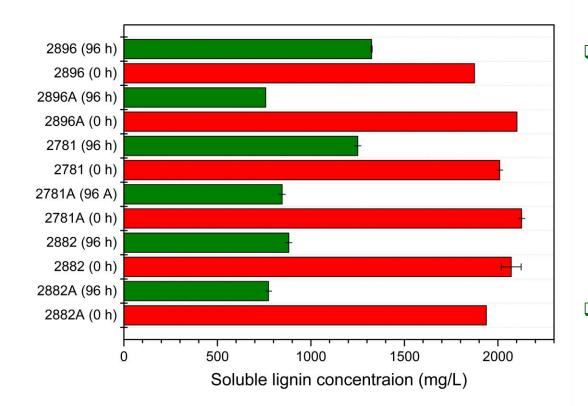


Fig. 6. Phenolics compounds degradation by adapted and non-adapted *R. toruloides* in hemicellulosic hydrolysate **Red:** Initial lignin soluble concentration Green: soluble lignin concentration at 96 h fermentation

- A decrease in phenolic compounds was observed in all tested conditions. A reduction in acid-soluble lignin was noted, being more expressive for the adapted strains of 63.93%, 60.11%, and 63.93% while for the non-adapted strains it was noted a reduction of 29.38%, 57.41%, and 37.77% for *R. toruloides* 2896, *R. toruloides* 2882 and *R. toruloides* 2781; respectively.
- These components are strong inhibitors of the fermentation process and, due to the variety available, it is difficult to determine the mechanism of inhibitory action provoked, since they can act alone or synergistically.

Table 2. Comparison of lipid production by R. to	oruloides in hemicellulosic hydrolysate
--	---

Yeast	pH _i	рН _f	Biomass (g.L ⁻¹)	Consumption of C (%)	Lipid accumulation % (m.m ⁻¹)	Yield biomass on carbohydrate Y _{x/s} (g.g ⁻¹)	Yield lipid on carbohydrate Y _{P/S} (g g⁻¹)	Lipid production rate QP (mg. L ⁻¹ .h ⁻¹)
Non- adapted								
2781	5.22	4.99	2.16	10.00	4.6%±1,04%	0,284	0.034±0,0077	1.0±0.2344
2882	5.22	7.30	9.45	24.00	3.5%±0.87%	1.126	0.048±0.0120	3.4±0,8520
2896	5.21	5.08	2.70	8.18	3.27%±0.17%	0.428	0.0377±0.2573	0.919±0,0486
Adapted								
2781	5.46	7.38	14.39	83	7.8%±0.22%	0.553	0.047±0.0017	11.6±0.42
2882	5,42	7.48	17.87	93.31	8.8%±0.68%	0.609	0.059±0.0045	16.3±1.26
2896	5.41	7.40	17.03	82	9.3%±0.36%	0.663	0.067±0.0026	16.6±0.63

**

□ The higher lipid accumulation was found in *R. toruloides* 2896 (9.3%), followed by *R. toruloides* 2882 (8.8%), and *R. toruloides* 2781 (7.8%) \Rightarrow evidencing the positive effect of cell-adaptation

Highlights

The adaptation process for strains of *R. toruloides* in non-detoxified sugarcane biomass hydrolysate was successful resulting in strains capable of growing and accumulating lipids in mixed sugars even in presence of microbial inhibitory compounds

□*R. toruloides* 2896 strain shows strong evidence that its lipid accumulation can be improved by optimizing the process conditions compared to the other strains. This finding is quite significant due to (1) high tolerance and degradation of inhibitory compounds present in the hydrolysate, (2) high accumulation of cellular biomass and (3) high yield of lipids

The production of lipids by *R. toruloides* is a promising alternative for the use of fermentable sugars from lignocellulosic biomass

ACKNOWLEDGEMENTS

FAPESP unesp

IPBEN

Instituto de Pesquisa em Bioenergia

Instituto de