



UPCYCLING MICROALGAE SIDE STREAMS FOR BIOPOLYMERS PRODUCTION

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INTRODUCTION



Nutrients



Treatment and/or valorization





Proteins

Carbohydrates

Pigments



INTRODUCTION

Polyhydroxyalkanoates (PHAs)
→ Bioco

Biocompatible, Biodegradable, Biopolyester

Similar properties to petrochemical plastics (e.g. melting temperature, flexibility, cristallinity)

Used as a substitute for conventional plastic

In line with the context of a circular economy, and EU guidelines for a sustainable bioeconomy for Europe



GOAL

PHA production process:

1. Fermentation of algae based side streams into volatile fatty acids (VFAs)

2. Enrichment of the mixed microbial culture (MMC) with the fermented stream and PHA accumulating organisms

3. PHA accumulation

Valorization of algae-based side streams into PHAs

Acidogenic fermentation batch tests





EXPERIMENTAL SET-UP: BATCH TESTS

Microalgae side streams (A4F – Algae for Future) +

Anaerobic sludge inoculum (Mutela WWTP, Portugal) \rightarrow

2 gVS/gVS (Substrate/inoculum)

Batch test conditions:

pH 5.5 30 °C 180 rpm 32 days 100 mL serum flasks Aerated with nitrogen gas

pH controlled with a phosphate buffer



EXPERIMENTAL SET-UP: BATCH TESTS



Nannochloropsis

Chlorella

Phototrophic consortium

Nannochloropsis Haematococcus

Ο

B

Organic load: 2.9 gC/L Strong buffer solution: 1 M Organic load: 1.2 gC/L Weak buffer solution: 0.1 M

Batch tests preformed in triplicate Control for each algae (no inoculum added) Control for inoculum (no microalgae added)

Gas phase analysed with GC Liquid phase analysed with HPLC





RESULTS- FEEDSTOCK CHARACTERIZATION

 Table 1: Characterization of each microalgae side stream

Feedstock	% Ash	% Protein	% Carbohydrates	% Lipids
<i>Nannochloropsis</i>	8.7 %	26.2 %	13.7 %	51.4 %
Chlorella —	10.4 %	22.8 %	25.2 %	41.6 %
Phototrophic consortium	9.9 %	25.9 %	30.3 %	33.9 %
Haematococcus	3.1 %	17.5 %	N/A	N/A
Note: N/A- Not Available				
*	,			

Rich in lipids

Rich in carbohydrates



RESULTS- BATCH TEST SET A



Figure 1: Results of fermentation products shown as mgCOD/L throughout the experiment. (A). Average results for triplicate experiment of *Nannochloropsis* (set A). (B) Average results for triplicate experiment of *Chlorella*. (C) Average results for triplicate experiment of phototrophic consortium.



RESULTS- BATCH TEST SET B



■ Lactate ■ Acetic acid ■ Propionic acid ■ Isobutyric acid ■ Ethanol ■ Butyric acid Ⅲ Isovaleric acid ■ Valeric acid ■ Caproic acid

Figure 2: Results of fermentation products shown as mgCOD/L throughout the experiment. (A). Average results for triplicate experiment of *Nannochloropsis* (set B). (B). Average results for triplicate experiment of *Haematococcus*.



RESULTS- BATCH TEST CONTROL



Figure 3: Results of fermentation products of controls shown as mgCOD/L throughout the experiment. (A). Results of *Nannochloropsis* control experiment, results of day 17 not available (set A). (B) Results of *Chlorella* control experiment. (C) Results of phototrophic consortium control experiment, results of day 14 not available. (D) Results of *Nannochloropsis* control experiment (set B). (E) Results of *Haematococcus* control experiment.



RESULTS- BATCH TESTS





SET A- Overall yield gC_{produced}/gC_{in}

Nannochloropsis: 17.7 % Chlorella: 37.7 % Phototrophic Consortium: 13.9% **SET B- Overall yield** $gC_{produced}/gC_{in}$

Nannochloropsis: 42.5 % Haematococcus: 21.0 %





CONCLUSIONS

Fermentation batch tests:

- > *Nannochloropsis* with low organic load showed highest yield of 42.5 % ($gC_{produced}/gC_{in}$)
- > Best production with the feedstock with lower content in carbohydrates and higher content in lipids
- Lower organic load and lower phosphate buffer solution favours a higher production and variety of VFAs
- Pre-treatments are being assessed



Thank you for your attention

ANY QUESTIONS?







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