## Preparation of xylo-oligosaccharides by deep eutectic solvent enhanced aqueous two-phase catalytic system

Yang Liu<sup>1</sup>, Can-Yang Shi<sup>1</sup>, Shuai You<sup>1, 2</sup>, Lu-Chan Gong<sup>1, 2</sup>, Jun Wang<sup>1, 2, \*</sup>

 <sup>1</sup> School of biotechnology, Jiangsu University of Science and Technology, 212018 Zhenjiang, China;
 <sup>2</sup> Sericultural Research Institute, Chinese Academy of Agricultural Sciences, 212018 Zhenjiang, China. Key words: Xylo-oligosaccharide; double aqueous phase; deep eutectic solvents; xylanase.

Presenting author email: wangjun@just.edu.cn

Presenting author email: wangjun(a)just.edu.c

Xylo-oligosaccharides (XOS) is a kind of oligosaccharide, with obvious benefits in reducing blood cholesterol, increasing calcium absorption, and maintaining gastrointestinal health and type II diabetes (Chen et al., 2016). At present, xylan-rich lignocelluloses are mainly prepared by chemical methods (alkali treatment, acid hydrolysis, hydrothermal method) and enzymatic hydrolysis (Liu et al., 2016). The acid hydrolysis is low in cost but difficult to separate the products generated by the reaction; the alkali treatment process requires a high concentration of alkali solution and high-temperature requirements (Singh et al., 2016). In comparison, the reaction process of the enzyme is mild, and no side reactions occur, which is suitable for practical application. Therefore, using an enzyme as a biocatalyst for the efficient production of oligosaccharides is a promising strategy.

Biphasic systems are systems made up of two immiscible components and are mainly used for the extraction and purification of downstream products, improving the current situation in homogeneous catalytic systems where the separation of product and catalyst is difficult, but not improving the environmental pollution caused by the use of organic solvents (Akama et al., 2002). The ATRP system, which uses water as the medium, has become a trend in recent years due to the low use of organic solvents, the mildness of the reaction steps and the environmental friendliness of the process. (Sun et al., 2018) The construction of aqueous catalytic systems for the reaction and separation of oligosaccharides can greatly improve the efficiency of industrial production and reduce the cost of post-reaction product separation (Chen et al., 2014). However, the construction of a biphasic system that separates the product oligosaccharides and the catalyst without destroying the xylanase activity is a difficult but highly relevant problem in practice.

Enzymes act as green catalysts in biochemical reactions and are able to speed up the reaction, therefore the selection of a suitable solvent is considered a key issue. Recently, the use of non-aqueous solvents has been used to replace aqueous solvents to facilitate the catalytic process of enzymes. DESs have attracted considerable interest as a green alternative to conventional solvents, not only because they are environmentally friendly, non-toxic and biodegradable organic compounds, but also their low cost, ease of production and several characteristics and properties (Harifi-Mood et al., 2018). Not only do DESs have specific properties compared to conventional organic solvents, but they are also non-destructive to enzyme activity. DESs have been reported to promote the catalytic efficiency and stability of laccase, cellulase and organophosphorus pesticide hydrolases. In the application of xylanase, there has been no research on enhancing enzyme stability and catalytic ability through deep eutectic solvents (Mann et al., 2009). Thus, the study of this aqueous two-phase catalytic system for strengthening xylanase by solvent system is very meaningful.

In this study, the relationship between different deep eutectic solvents and the catalytic capacity of xylanase was investigated and the deep eutectic solvent with the best effect was screened. On this basis, a deep eutectic solvent with salt and PEG catalytic separation system was constructed to prepare oligosaccharides.



Figure 1 Effect of deep eutectic solvents on xylanase, (A) Catalytic capacity; (B) Temperature stability at 90 °C.

Fig 1A shows deferent DES different compositions of DES had different effects on the catalytic capacity of xylanase, with UBH and MBH having a significant activity destruction of xylanase, while several other DES had an enzymatic activity enhancing effect on xylanase. Gly2B prepared by betaine and glycerol was the most effective, and the xylanase activity could be increased by 53.24% at 30% concentration of Gly2B. Fig 1B shows the different DES all improve the stability of xylanase and the hydrophobic environment better protects the active centre of xylanase, with betaine and sorbitol as hydrogen acceptors for SoBH and Gly2B being the most effective.

Table. I Killette constants of Xylandse in different solvents.				
Solvent	$K_{ m m}$	$k_{ m cat}$	$V_{\max}$	$k_{\rm cat}/K_{\rm m}$
	(mg/mL)	(/s)	(U/mg)	(mL/s/mg)
Buffer	3.69±0.13 <sup>a</sup>	2530.28±273.49 <sup>a</sup>	4804.33±519.29 °	693.66±184 <sup>a</sup>
Gly2B	3.32±0.16 <sup>b</sup>	3105.53±143 <sup>b</sup>	5896.57±374 <sup>b</sup>	933.97±104 <sup>b</sup>

Table.1 Kinetic constants of xylanase in different solvents

Table 1 shows that In Gly2B, the *K*m of xylanase was 13.45% lower and the catalytic efficiency was 24.63% higher than in the buffer. Therefore, the substrate affinity of xylanase is increased in the deep eutectic solvent, allowing for better catalysis.

In conclusion, deep eutectic solvents with different compositions were prepared, and the catalytic ability and stability of xylanase in different solutions with different concentrations were analyzed. On this basis, an aqueous two-phase catalytic system was constructed to prepare and separate xylo-oligosaccharides.

Acknowledgements: This work was supported by National Natural Science Foundation of China (grant number 21978121), Natural Science Foundation of Jiangsu Province (BK20190957), 333 High-level Talent Training Project of Jiangsu Province (BRA2019281).

## **References:**

- [1] Chen, X. Q., Yang, Q. L., Si, C. L., Wang, Z. J., Huo, D., Hong, Y. M., Li, Z. Q., 2016. 'Recovery of oligosaccharides from prehydrolysis liquors of poplar by microfiltration/ultrafiltration membranes and anion exchange resin'. *REVIEWS IN CHEMICAL ENGINEERING*. 4 (3): 937-943.
- [2] Liu, X., Meng, X. Y., Xu, Y., Dong, T., Zhang, D.Y., Guan, H. X., Zhang, Y., Wang, J., 2019. 'Enzymatic synthesis of 1-caffeoylglycerol with deep eutectic solvent under continuous microflow conditions'. *BIOCHEMICAL ENGINEERING JOURNAL* 142 (2): 41-49.
- [3] Singh, R. D., Banerjee, J., Sasmal, S., Muir, J., Arora, A., 2018. 'High xylan recovery using two stage alkali process from high lignin biomass and its valorisation to xylooligosaccharides of low degree of polymerisation'. *BIORESOURCE TECHNOLOGY*. 256 (8): 110-117.
- [4] Akama, Y., Sali, A., 2002. 'Extraction mechanism of Cr(VI) on the aqueous two-phase system of tetrabutylammonium bromide and (NH4)2SO4 mixture'. *TALANTA*. 57 (4): 681-686.
- [5] Sun, Z., Glebe, U., Charan, H., Böker, A. and Wu, C., 2018. 'Enzyme–Polymer Conjugates as Robust Pickering Interfacial Biocatalysts for Efficient Biotransformations and One-Pot Cascade Reactions'. ANGEWANDTE CHEMIE INTERNATIONAL EDITION, 57 (42): 13810-13814.
- [6] Chen, J., Wang, Y., Ding, X., Huang, Y., Xu, K., 2014. 'Magnetic solid-phase extraction of proteins based on hydroxy functional ionic liquid-modified magnetic nanoparticles'. ANALYTICAL METHODS, 6(20), 8358-8367.
- [7] Harifi-Mood, A. R., Sadrzadeh, S., 2018. 'Dimethyl sulfoxide/deep eutectic solvents mixtures as media in the reaction of 1-fluoro4 initrobenzene with piperidine: a solvent effect study'. JOURNAL OF PHYSICAL ORGANIC CHEMISTRY, 31 (4): e3787.
- [8] Mann, J. P., Mccluskey, A., Atkin, R., 2009. 'Activity and thermal stability of lysozyme in alkylammonium formate ionic liquids-influence of cation modification'. *GREEN CHEMISTRY*. 11 (6): 785-792.