

EVALUATION OF THE USE OF FOOD WASTE-DERIVED VOLTATILE FATTY ACIDS EFFLUENT IN POLYHYDROXYALKANOATES PRODUCTION BY *CUPRIAVIDUS*NECATOR DSM 545

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1. Introduction

Starch and sugar-based substrates have been the main feedstock in polyhydroxyalkanoates (PHAs) production. However, the use of those feedstock accounts for a significant share of up to 30-50% in the total production cost, which highly hinders the PHA commercialization. In this regard, fermentable compounds such as volatile fatty acids (VFAs) have emerged as promising alternatives to the expensive conventional feedstock in PHA production. VFAs not only directly engage in PHA biosynthesis but also can be produced in large amount via anaerobic digestion of various organic wastes. Therefore, the aim of this study was to employ food waste-based VFA stream as the economical feedstock for PHA production. The application of VFAs in PHA-bearing microbial cultivation is believed to increase the cost-competitiveness of PHA production and the sufficiency in resource uses.

2. Material and Methods

Synthetic VFAs with different loadings were initially employed to determine the feasible VFA concentrations for the growth of *Cupriavidus necator* DSM 545. Afterwards, a mixture of synthetic VFAs mimicked real VFA effluent of 4.45 g/L was employed to evaluate the bacterial growth in VFA mixture. Lastly, the cultivation of *C. necator* was conducted using real VFA effluent and without any nutrient supplement. All experiments were executed in duplicate using a 2-L continuous stirred tank reactor (CSTR) at 32°C, 120 rpm and pH control at 7 using 2M NaOH and 2M HCl. In prior to each cultivations, the media preparations were performed in sterilized conditions.

The VFA effluents used in this study were produced and recovered from acidogenic fermentation of food waste provided by Wainaina et al. (2020). Cell density, substrate concentration, biomass and PHA accumulation were measured at different interval times to assess the fermentation performance.

3. Results and Discussion

As presented in Figure 1a, acetic and butyric acid were found to be favourable carbon sources for the cultivation of *C. necator*, the concentrations of which were almost depleted after 36h. Whilst the assimilation of propionic and isobutyric acid was slower at the same concentration of 5 g/L. The concentration of valeric, isovaleric and caproic acids, on the other hand, was diluted to 2.5 g/L to achieve the feasible bacterial growth. The changes in cell density were measured throughout the fermentation, showing the similar pattern corresponding to the VFA consumption (Figure 1b). Regardless the potential in replacing conventional substrates in PHA production, the use of VFAs could pose a drawback of substrate inhibition at excessive concentration of above 5 g/L (Agustín Martinez et al. 2015; Yun et al. 2013).

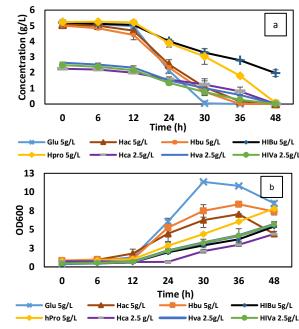


Figure 1. Changes in cell density (a) and volatile fatty acid (b) concentration during the fermentation in a bioreactor using individual VFAs at different concentrations.

A significant change in metabolic behavior was observed as the bacteria was cultivated in a synthetic VFA mixture (Figure 2a). In this regard, butyrate acid became the most preferable component which was consumed the first at an accelerating rate among the blend of VFAs. The consumption of acetic acid, on the other hand, was only conducted after the depletion of other VFAs. The similar phenomenon was observed in the case of food waste-based VFAs as the substrates were assimilated in the same pattern but at a faster uptake rate (Figure 3a).

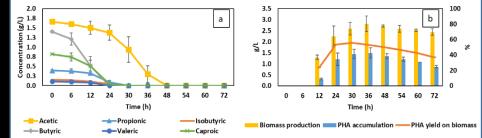


Figure 2. Changes in volatile fatty acid (VFA) concentration (a), and biomass and PHA production (b) using synthetic mixture VFAs during the fermentation in a bioreactor.

However, the biomass and PHA yield obtained in synthetic VFAs were higher than that of acquired in real VFA effluent, 2.8 and 1.49 g/L compared to 2.26 and 1.02 g/L, respectively (Figure 2b & 3b). Generally, the PHA yield obtained in this study was notably higher than those reported in other studies by Cerrone et al. (2014), Tajima et al. (2003), (Agustín Martinez et al. 2015).

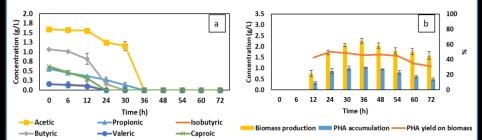


Figure 3. Changes in volatile fatty acid (VFA) concentration (a), and biomass and PHA production (b) using food waste-based VFA during the fermentation in a bioreactor.

Nuclear Magnetic Resonance (NMR) analysis was used to determine the composition and crystallinity of the accumulated PHAs (Table 1). The PHA obtained was found to be a copolymer of poly(3-hydroxybutyrate-co-3-hydroxyvalerate) (PHBV) and the crystallinities of which were in compliance with samples obtained from a study by William et al. (1995).

Table 1. Composition and total crystallinity of the copolymers produced from two different carbon sources.

Sample	Fraction %		TD 4.1 4.191.14
	НВ	HV	Total crystallinity
Synthetic mixture VFAs	93.8	6.2	62.2
Food waste-based VFAs	93.5	6.5	61.4

4. Conclusion

In this study, food waste-derived VFAs can be used directly for the cultivation of PHA-bearing bacteria without the addition of external nutrients. In this regard, a maximum biomass production of 2.26 g/L was obtained, corresponding to the PHA content of 1.02 g/L. The PHA obtained was PHBV with the HV content of 6.2%. The use of organic waste-based VFAs provides a great opportunity to replace the expensive conventional feedstock, increasing the cost-competitiveness of PHA production.

5. References

- Agustín Martinez, G., Bertin, L., Scoma, A., Rebecchi, S., Braunegg, G. & Fava, F. (2015). Production of
 polyhydroxyalkanoates from dephenolised and fermented olive mill wastewaters by employing a pure
 culture of Cupriavidus necator. Biochemical Engineering Journal, 97, pp. 92-100.
- Cerrone, F. et al. (2014). Medium chain length polyhydroxyalkanoate (mcl-PHA) production from volatile fatty acids derived from the anaerobic digestion of grass. Appl Microbiol Biotechnol, 98(2), pp. 611-20.
 Tajima, K., Igari, T., Nishimura, D., Nakamura, M., Satoh, Y. & Munekata, M. (2003). Isolation and
- Tajima, K., Igari, T., Nishimura, D., Nakamura, M., Satoh, Y. & Munekata, M. (2003). Isolation and characterization of Bacillus sp. INT005 accumulating polyhydroxyalkanoate (PHA) from gas field soil. J Biosci Bioeng, 95(1), pp. 77-81.
- Wainaina, S., Awasthi, M. K., Horváth, I. S. & Taherzadeh, M. J. (2020). Anaerobic digestion of food waste to volatile fatty acids and hydrogen at high organic loading rates in immersed membrane bioreactors. Renewable Energy, 152, pp. 1140-1148.
- William, J. O., David, L. V., Terry, L. B. & Robert, H. M. (1995). Cocrystallization in random copolymers of poly(β-hydroxybutyrate-co-β-hydroxyvalerate) and its effect on crystalline morphology. Canadian Journal of Chemistry, 73(11), pp. 2094-2100.
- Yun, J., Sawant, S. & Kim, B. S. (2013). Production of Polyhydroxyalkanoates by Ralstonia Eutropha from Volatile Fatty Acids. Korean Journal of Chemical Engineering, 30.